

Characteristics and electrochemical membrane sensors for selective study and determination of some antidepressant drugs in their pharmaceutical preparations

Anwar A. Wassel^{1,*} and Shabaan A. Abdullatif²

¹ Department of Chemistry, Faculty of Science, Imam Abdulrahman Bin Fasil University –KSA.

² National Organizations for Drug Control and Research. (NODCAR) Egypt.

World Journal of Advanced Research and Reviews, 2023, 20(02), 804–819

Publication history: Received on 29 September 2023; revised on 12 November 2023; accepted on 14 November 2023

Article DOI: <https://doi.org/10.30574/wjarr.2023.20.2.2294>

Abstract

The construction and electrochemical characterisation of potentiometric sensors of the PVC-membrane type are described being developed for the determination of selected antidepressants; namely: *Olanzapine*, *Oxazepam*, and *Lorazepam*. The sensing PVC-membranes incorporate the ion-associates of the drug cations with ammonium reineckate as selective materials dispersed in 2-nitrophenyl-octylether (NPOE) or dibutyl sebacate (DBS) as the plasticizers of choice. These ISEs exhibit rapid, stable and nearly Nernstian response over a relatively wide concentration range of $1 \times 10^{-7} \text{ M} - 0.01 \text{ mol L}^{-1}$ for *Olanzapine* and $1 \times 10^{-6} - 0.01 \text{ mol L}^{-1}$ for *Oxazepam* and *Lorazepam*, in mild acidic solutions with pH 4.5. No interferences caused by either inorganic or organic species were found. The three electrodes developed have been tested for potentiometric titrations of all three drugs, as well as for their direct determination via the respective calibration curves. The individual experiments then resulted in average recovery rates in an interval of 98.4–99.2 %, with the R.S.D. of *ca.* $\pm 1.5 \%$ ($n = 3$) for all three analytes of interest. Finally, the method proposed has been applied to the determination of the three drugs in pharmaceutical formulations and in urine, when the results obtained were comparable to those obtained with the reference HPLC measurements.

Keywords: Potentiometry; PVC-membrane ion-selective electrodes; Ammonium reineckate; Antidepressant drugs; Pharmaceutical formulations; Urine.

1. Introduction

Benzodiazepine drugs (BZD) belong to a group of substances that are known for their sedative, hypnotic, tranquillizing, and anticonvulsant properties and are being prescribed worldwide as therapeutics to treat anxiety, sleep disorders, status epileptics, insomnia, frequent nocturnal awakenings, early morning awakenings schizophrenia, and related disorders. In addition, they are used as muscle relaxants for alleviation of panic attacks and as induction agents in anesthesiology [1].

Olanzapine ("Olan"; see Figure 1) is an antipsychotropic agent that belongs to the thienobenzo-diazepine class, a relatively new benzodiazepine which has been found useful to treat schizophrenia and some other psychosis [2-4]. *Olanzapine* is a crystalline solid substance of yellow colour and with a melting point of 263.2°C [5]. Its molecular formula is $\text{C}_{17}\text{H}_{20}\text{N}_4\text{S}$, corresponding to a molecular weight of 312.44 g mol⁻¹.

* Corresponding author: Anwar A. Wassel.

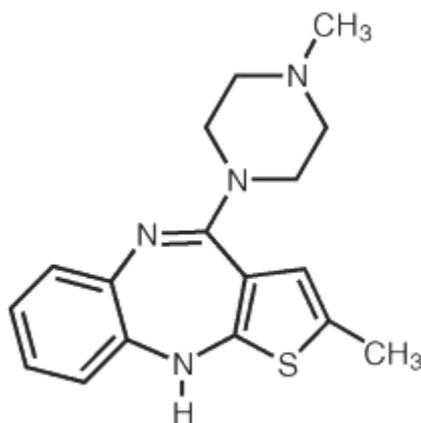


Figure 1 Structural chemical formula of *Olanzapine* / Methyl-4-(4-methyl-1-piperazinyl)-10*H*-thieno- [2,3-*b*] [1,5]benzodiazepine.

Oxazepam (aka *Oxazem* or *Serax*-tablets, "*Oxaz*"; Figure 2) is an anti-anxiety agent of the benzodiazepine type used primarily for treating mild to moderate anxiety. It is also prescribed as a sedative, hypnotic, anticonvulsant, and muscle relaxant. [6-9].

This drug is frequently encountered in clinical and forensic toxicology, having been featured in an increasing number of misuses and abuses over the past years [10, 11]. Finally, it is also used to treat irritable bowel syndrome. *Oxazepam* is a creamy white or pale yellow crystalline substance, odourless, and with melting point of 205°C. Its molecular formula is C₁₆H₁₃ClN₂O for both possible enantiomers, corresponding to a molecular weight of 284 g mol⁻¹.

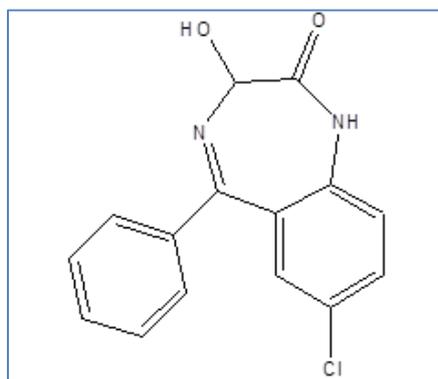


Figure 2 Chemical formula of *Oxazepam* / 7-Chloro-2,3-dihydro-3-hydroxy-5-phenyl-1*H*-1,4-benzo-diazepin-2-one.

Lorazepam ("*Lora*"; Figure 3) is classified pharmacologically as a short acting benzodiazepine and under the trademark *Ativan*[®] (1mg or 2mg of active substance) is a generic representative for the popular anti-anxiety medication, indicated for generalized anxiety disorder (GAD), panic disorder, anxiety associated with depression and insomnia. Also, it is sometimes used as a sleeping agent, anti-convulsant, and for alcohol detoxification [12-15]. *Lorazepam* / *Ativan*[®] is a white — or, almost white — microcrystalline powder with a melting point of 245 °C. Its molecular formula is C₁₅H₁₀Cl₂N₂O₂, corresponding to a molecular weight of 321.2 g mol⁻¹.

In this article, a simple and rapid method for the determination of the three, above described pharmaceuticals has been developed based on the employment of ion-selective electrodes (ISEs) whose PVC-membranes are saturated with the corresponding BZD / reineckate ion-associate and that can be used for indication in direct potentiometry, as well as in potentiometric titrations.

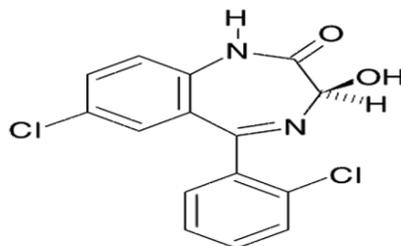


Figure 3 Chemical formula of *Lorazepam* / 7-Chloro-5-(2-chlorophenyl)-1,3-dihydro-3-*1H*-1,4-benzo-diazepine-2-one.

2. Experimental Method

2.1. Chemicals and Reagents

All reagents and chemicals used throughout the work were of analytical-reagent grade and solutions were prepared with deionised distilled water (DDW). Both polyvinylchloride (PVC) powder and ammonium reineckate (A-RE; purity: 98 %) used as reagent and titrant were obtained from Aldrich. Dibutyl sebacate (DBS; a plasticizer of purity ~ 99%) was purchased from BDH (British Drug Houses, Pool, England); 2-Nitrophenyl octyl ether (o-NPOE; a plasticizer of purity ca. 99 %) from Fluka. Tetrahydrofuran (THF; a solvent with purity of about 99%) was purchased from Aldrich.

An acetate buffer solution of pH 4.5 was prepared by dissolving 13.6 mg sodium acetate with approx. 5 mL glacial acetic acid in 1000 mL deionised and doubly distilled water. A phosphate buffer solution of pH 4.5 was prepared by mixing the appropriate quantities of 0.1 M H_3PO_4 with 0.2 M Na_2HPO_4 in 500 mL deionised distilled water.

In interference studies, selected inorganic cations, amino acids, and sugars were examined via 1×10^{-3} M solutions prepared in acetate buffer (pH 4.5), when containing the following species: Na^+ , K^+ , Mg^{2+} , Ca^{2+} , Sr^{2+} , Cu^{2+} , Zn^{2+} , and diphenhydramine. The remaining solutions of urea, starch, glucose, and maltose were 0.01 mol L^{-1} .

2.2. Standard Solutions and Precipitates for Membrane Modification

Standard Solutions of the Drugs: The stock solutions of *Olanzapine*, *Oxazepam*, and *Lorazepam* (or *Olan*, *Oxaz*, and *Lora*, resp.) were prepared as 0.01 mol L^{-1} , by dissolving the proper weight of solid substance into suitable amount of 0.002 M HCl added in droplets under continuous stirring till a complete dissolution of the drug was achieved.

The resultant solution was made up to 100 mL in a measuring flask using de-ionized water. Working diluted standards were then prepared in a series of calibration solutions in the concentration range from 1×10^{-7} up to 0.01 mol L^{-1} for *Olan* and from 1×10^{-6} to 0.01 mol L^{-1} for *Oxaz* or *Lora*; all being made as solutions in acetate buffer (pH 4.5). Both stock solutions and diluted standards were kept in dark and in a refrigerator (at 4°C).

Standard of ammonium reineckate; i.e., ammonium tetrathiocyanato-diammine chromate(III), $\text{NH}_4[\text{Cr}^{\text{III}}(\text{SCN})_4(\text{NH}_3)_2] \times \text{H}_2\text{O}$; (A-RE): A 0.01 M standard solution was prepared by dissolving 2.6300 g in 100 mL of deionised distilled water.

Preparation of the BZD / Reineckate Ion-Associates: Each BZD / RE ion-pair complex was prepared by mixing the equal volumes of 0.01 M *AmRe* with 0.01 M *Olan*, *Oxaz* or *Lora*; when the latter being added drop-wise under continuous stirring. The resultant precipitates were left in contact with their native liquor overnight to ensure the complete coagulation, then filtered off through a *Whatman*[®] filter paper No. 42, washed thoroughly and repeatedly with distilled water, and being left to dry at room temperature for at least 24 hours and finally grinded to fine powder.

2.3. Apparatus and Other Equipment

All potentiometric measurements were made at $25 \pm 2^\circ\text{C}$ using a pH meter (model "H-8417", Hanna, further specification, see Ref.[16]) with pH sensitivity of ± 0.05 pH units with Drug-PVC matrix membrane sensors in conjunction with a double-junction Ag/AgCl reference electrode (model "900200", Orion) containing a solution of (10%, w/w) KNO_3 in the outer compartment. The combined glass electrode (model "F-22E"; Horiba, [16]) was used for all pH measurements after calibration with the aid of a set of commercially available buffers.

The electrochemical system used can be represented as follows: Ag | AgCl | KCl (saturated, *ca.* 3 mol·L⁻¹) || Salt bridge || Sample, it means: Reference electrode half-cell | Membrane | Filling solution | Ag/AgCl | Ion-selective electrode half-cell; where the filling solution is a mixture of 0.01 M solution for each of investigated drugs and KCl.

2.4. Procedures and Samples

Membrane Preparation: A 10 mg portion of drug / reineckate ion-pair complex was thoroughly mixed in a glass Petri dish (5 cm in diameter) with 190 mg of PVC powder and 350 mg of 2-nitrophenyl octylether (NPOE) or dibutyl sebacate (DBS).

The resultant cocktail was dissolved in 5 mL THF, covered with a filter paper and left overnight to evaporate to dryness at ambient temperature. Using this procedure, a semi-transparent and plastic-like membrane with a thickness 0.1 mm could be obtained. Then, the parts of the membrane material were cut using a cork borer and, when plasticizing with THF, an exchangeable PVC-tip was clipped onto the end of the electrode glass body. The tip was left to stand for 24 hours to allow complete evaporation of THF and, finally, the fabricated electrode was soaked into 0.01 M solution of the corresponding drug for 24 hours before use (and also being stored in the same solution when not in use).

The constructed sensor was washed with deionised distilled water and blotted with tissue paper between measurements. The effect of membrane composition was studied for Olanzapine-modified sensor which was plasticized with NPOE — by weighing three different amounts of ionophore (5 mg, 10 mg, or 15 mg, respectively), and mixing with 190 mg PVC + 350 mg NPOE + 5 mL THF in a glass mortar. To obtain homogeneous and uniform thickness of the mixture [17], the membranes were left to dry free (not less than 24 hours) and, during later evaluation, plotted as a function of *Olan* ion concentration.

Measurement and Construction of Calibration Curves: 10 mL aliquots of each sample solutions – in acetate buffer of pH 4.5 – having concentrations of 1×10^{-7} – 0.01 M for *Olan* or 1×10^{-6} – 0.01 M for *Oxaz* and *Lora* were transferred into 50mL beakers. The potential in mV of each sample solution was directly measured using the respective (drug / reineckate ion-pair modified) ISEs based on DBS or NPOE as plasticizer as mentioned in the previous section.

The solutions were stirred and the potential readings were recorded from the low to the high concentration after stabilization to ± 0.3 mV [18]. The potentials were plotted as a function of $-\log$ (drug concentration). These graphs were used for the subsequent determination of unknown concentrations of corresponding drug. The theoretical (aka “lower”) detection limit was determined at the point of intersection of the extrapolated linear segments of the corresponding drug calibration curve.

Studies on pH-Effect and Response Time: The effect of pH of the test solution on the potential values of the electrode system in solutions of different concentrations of pharmaceutical compounds was tested using two concentrations of each drug ion (1×10^{-3} and 1×10^{-4} M) over a range of pH (2-11) and recording the potential readings of the studied sensors by immersing the Ross combination glass electrode, corresponding drug-sensor and a double junction Ag/AgCl reference electrode in 50 mL beakers containing 25 mL aliquots of 10^{-3} and/or 1×10^{-4} M drug-aqueous solutions. The pH of each solution was gradually changed by adding small aliquots of dilute sodium hydroxide and/or hydrochloric acid solutions. The mV-vs.-pH profile of each drug concentration was plotted for each electrode system. The potential readings that were insensitive to pH changes were obtained from the mV-pH plots. The dynamic response time of the ISEs could also be characterised by the fact that the respective measurements required to reach a steady-state potential within ± 0.3 mV only, regardless that analysed concentration differed also in one order.

Determination of Sensor Selectivity: The influence of some inorganic cations, sugars and amino acids, used as additives or binders, on the response of the electrodes towards their respective drugs was in tested. The separate solution method [19], which requires two potential measurements, first, the potential is measured in a solution containing a known activity of the ion for which the electrode is selective, second, the potential is measured in a solution containing the interfering ion, was used to determine the selectivity coefficient value, $K_{d,i}$; see later in Table 3. Separate drug primary ion (d) and interfering secondary ion (i) solutions having equal concentrations were prepared. Their potentials E_d and E_i were measured using prepared ISEs in the previous mentioned cell. Selectivity coefficients were calculated using either one of the following equations:

$$\text{Log } K_{d,i} = (E_i - E_d) / S \quad (1a)$$

$$\text{Log } K_{d,i} = (E_i - E_d) / S + (1 - z_d / z_i) \log [d] \quad (1b)$$

where "E_i" and "E_d" are the electrode potentials of 0.01 M solution of each of the investigated drug and interfering cation, I, respectively. Equation (1a) is used for mono-valent secondary ions whereas equation (1b) is used for divalent or higher ones. "z_d", "z_i" are the charges on the primary or secondary ions, and "S" is the slope of calibration curve for the primary ion.

The individual interfering inorganic ions and organic species used in the studies on the selectivity of the different membranes were already specified above. For the proper measurements, the respective drug solution was transferred into a 50 mL beaker containing 9.0 mL acetate buffer (pH 4.5). The drug sensor in conjunction with a double junction Ag/AgCl reference electrode was immersed into the solution and the potential read (in mV). Then, the solution with interfering species was transferred into a 50 mL beaker containing 9.0 mL the same buffer and the potential change was again recorded (as E_i). Selectivity coefficients were calculated as shown before.

Potentiometric Titrations. The individual aliquots of 0.01 M BZD were transferred into 50 mL titration cell and diluted to 10 mL by acetate buffer of pH 4.5. The resulting solutions were stirred and titrated against 0.01 M A-RE according to the corresponding ion-exchanger used in constructing the electrode. The proper titration curves; i.e., the electrode potential vs. the titrant volume (E-vs.-V) plots and their end-points were evaluated conventionally.

Pharmaceutical Samples of the Drugs Studied. Five tablets of the drug formulations were weighted and finely powdered in a small mortar. An accurately weighted portion of powdered drug, equivalent to 1.562 mg *Olan*, 1.433 mg *Oxaz*, or 1.606 mg *Lora*, was transferred into 5 mL measuring flask dissolved in the minimum volume of de-ionized distilled water and few drops of 0.002 M HCl. The solution was filtered into a 50mL calibrated flask, then diluted to the mark with acetate buffer of pH 4.5 and shaken well.

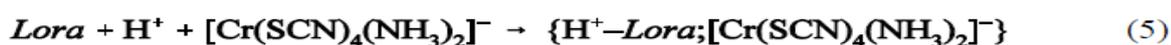
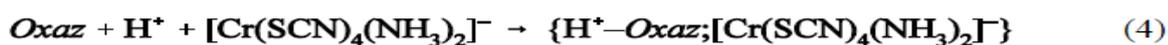
A series of 10, 15, and 20 mL aliquots of each drug solution were transferred into 25 mL volumetric flasks and made up to the mark with acetate buffer. Thus, solutions having concentrations of 12.5, 18.75 and 25 µg·mL⁻¹ *Olan*, 11.46, 17.2 and 22.92 µg·mL⁻¹ *Oxaz*, or 12.85, 19.30 and 29.70 µg·mL⁻¹ *Lora* were made and the potentials of the resultant drug solutions could be directly measured at the corresponding ISE.

Biological Fluids (Drugs in Urine): Solutions of 5×10⁻³ M *Olan* (31.24 µg·mL⁻¹), *Oxaz* (28.66 µg·mL⁻¹) or *Lora* (32.12 µg·mL⁻¹), were prepared by dissolving the appropriate amount of pure substances — specifically: 1.562 mg, 1.433 mg, and 1.606 mg — in 50 mL in the order and using solvent as described above. Aliquots of 10, 15, and 20 mL (pipetted from the respective drug solutions) were transferred into 25 mL measuring flasks; each containing 5 mL urine, and filled up to the mark with solvent. The actual electrode potentials of the individual solutions were measured in the same way as above.

3. Results and Discussion

Principles of the Electrode Functioning

According to the literature (see, e.g., Ref. [18]), the respective drug moiety can be protonised and such a cation can readily react with the anionic form of reineckate, following the scheme(s)



Where H⁺-(OLAN)₂ or H⁺-OXAZ and H⁺-LORA, respectively, denote the protonated form of the respective drug and the compounds on the right-hand side the resultant ion-associates.

3.1. Three Electrodes Studied and Their Operability

Olan-, *Oxaz*-, and *Lora*-ISEs with the electroactive material chosen; i.e., drug / reineckate were constructed using PVC as a matrix and NPOE or DBS as the plasticizers. The ISEs were investigated in order to compare their electrode performance and the results are summarized in Tables 1 and 2. The corresponding calibration graphs, given in Figs. 4, 5 and 6, indicate that for all electrodes tested, having nearly Nernstian cationic response slopes (26.12 and 24.22

mV/decade with a limit of detection 1.2×10^{-7} and 0.9×10^{-7} M for *Olan*-ISE, plasticized with NPOE or DBS, respectively, and 52.10 – 54.30 mV/decade for both *Oxaz*-ISE and *Lora*-ISE.

Table 1 Potentiometric response characteristics for *Olan-RE* / membrane ISE with NPOE or DBS as plasticizer.

Parameter evaluated	Olanzapine-RE sensor	
	NPOE	DBS
Concentration range [mol L ⁻¹]	10^{-7} - 10^{-2}	10^{-7} - 10^{-2}
Slope [mV / decade]	26.12	24.22
Intercept	162.77	168.7
R.S.D [% , ±]	0.62	0.77
Limit of detection [mol L ⁻¹]	3.2×10^{-7}	4.2×10^{-7}
Correlation coefficient (r ²)	0.9939	0.9933
Actual pH (range)	4.5	4.5
Response time [s]	20	20
Life span [weeks]	4	4

Table 2 Potentiometric response characteristics for *Oxaz*- and *Lora*-membrane ISEs with NPOE or DBS as plasticizer

Parameter evaluated	Oxa-RE sensor		Lora-RE sensor	
	NPOE	DBS	NPOE	DBS
Concentration range [mol L ⁻¹]	10^{-6} - 10^{-2}	10^{-6} - 10^{-2}	10^{-6} - 10^{-2}	10^{-6} - 10^{-2}
Slope [mV / decade]	54.30	52.10	55.01	54.43
Intercept	354.82	355.36	343.88	354.53
Limit of detection [mol L ⁻¹]	4.6×10^{-6}	5.3×10^{-6}	3.7×10^{-6}	4.8×10^{-6}
R.S.D [% , ±]	0.56	1.20	0.74	1.30
Correlation coeff. (r ²)	0.9989	0.9978	0.9979	0.9978
Actual pH (range)	4.5	4.5	4.5	4.5
Response time [s]	30	30	40	40
Life span [weeks]	6 – 8	6 – 8	6 – 8	6 – 8

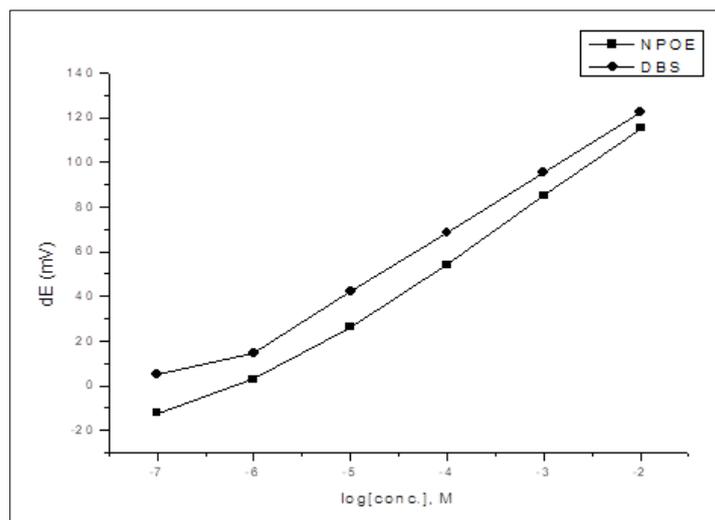


Figure 4 Calibration curve of *Olan*-membrane ISEs with NPOE or DBS as plasticizer.

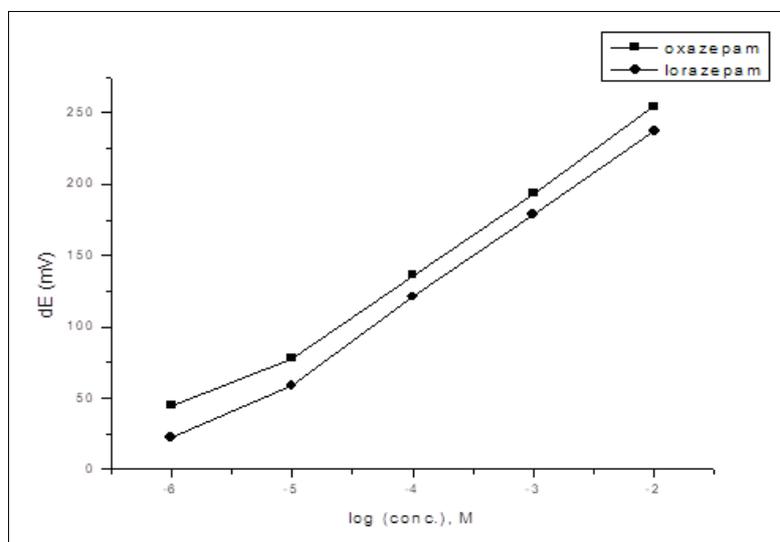


Figure 5 Calibration curve of *Oxaz*- and *Lora*-membrane ISEs with NPOE as plasticizer.

The linearity range extended down to 1×10^{-7} mol L⁻¹ for *Olan* and 1×10^{-6} mol L⁻¹ for *Oxaz* or *Lora*. Also, detection limits that were estimated according to a widely adopted criterion “three sigma” [19, 20], could be evaluated from the calibration curves. Although the three different electrodes had exhibited certain differences between linearity ranges and response slopes, the resultant nuances were found insignificant.

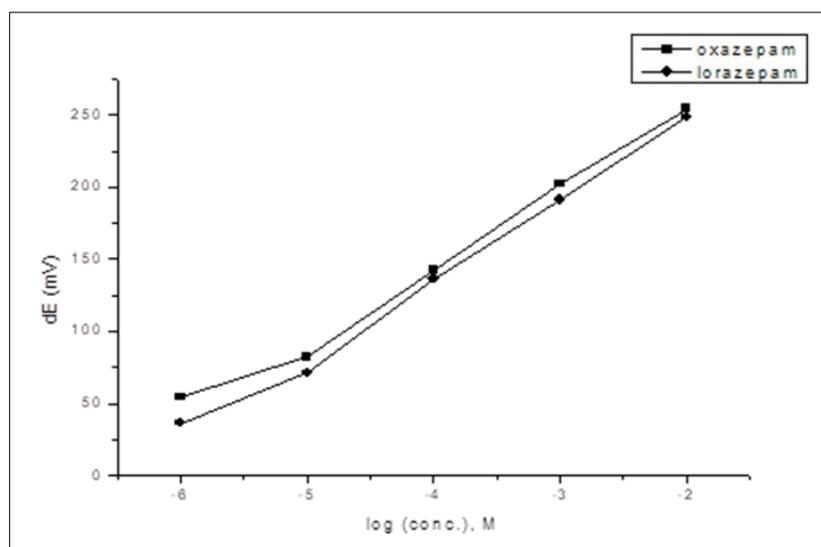


Figure 6 Calibration curve of Oxaz- and Lora-membrane ISEs with DBS as plasticizer

3.2. The Electrodes and Their Selectivity Characteristics

The selectivity coefficient is the main source of information concerning interferences on the electrode response. In analytical applications, the selectivity for the analyte must be as high as possible, i.e. the selectivity coefficients must be very small, so that, the electrode exhibits a Nernstian dependence on the primary ion over a wide concentration range. The selectivity of the ion-exchanger of membrane electrodes depends on the selectivity of the ion-exchange process at the membrane test solution interface and the mobility of the respective ions in the matrix of the membrane. The hydrophobic interactions between the primary ions and the PVC membrane are reflected by the values of the Gibb's energy of transfer for cations between the aqueous and membrane phases. The response of the electrodes towards different substances and ionic species was checked.

Table-3 Potentiometric selectivity coefficients, K_{ij}^{pot} , for the three proposed electrodes based on NPOE [where (i) denotes mediator and (j) the interfering species].

Interfering species (ion / substance)	Olan-sensor	Oxa-sensor	Lora-sensor
	K_{ij}^{pot}	K_{ij}^{pot}	K_{ij}^{pot}
Na ⁺	8.5×10^{-4}	1.6×10^{-3}	7.2×10^{-4}
K ⁺	7.7×10^{-4}	4.4×10^{-3}	5.6×10^{-3}
Mg ²⁺	5.3×10^{-4}	5.8×10^{-3}	4.1×10^{-3}
Ca ²⁺	8.5×10^{-3}	7.3×10^{-4}	6.3×10^{-4}
Sr ²⁺	1.2×10^{-2}	6.6×10^{-3}	3.3×10^{-3}
Cu ²⁺	6.3×10^{-2}	5.2×10^{-3}	6.1×10^{-3}
Zn ²⁺	2.3×10^{-2}	8.7×10^{-3}	4.3×10^{-3}
Diphenhydramine	3.4×10^{-3}	2.3×10^{-4}	1.2×10^{-4}
Urea	8.2×10^{-4}	2.2×10^{-4}	2.4×10^{-4}
Starch	2.4×10^{-4}	4.3×10^{-4}	8.1×10^{-4}
Glucose	3.6×10^{-4}	3.6×10^{-4}	7.4×10^{-4}
Maltose	5.4×10^{-4}	6.2×10^{-4}	6.6×10^{-4}
Olanzapine	-	0.7×10^{-2}	3.2×10^{-2}
Oxazepam	3.2×10^{-2}	-	3.1×10^{-2}
Lorazepam	1.3×10^{-2}	4.2×10^{-2}	-

As mentioned in the Experimental part, the selectivity coefficients of the interfering cations were determined by the separate solution method which is a very simple approach, providing a reasonable measure of the degree of interference of foreign species that may be present in the test solution [21, 22]; see Table III. The mechanism of selectivity is mainly based on the stereo-specificity and electrostatic environment and it is dependent on how much fitting is present between the locations of the lipophilicity sites in the two competing species in the drug solution side and those present in the receptor of the ion-exchanger.

The inorganic cations do not interfere because of differences in ionic size, mobility and permeability. Also, the smaller the energy of hydration of the cation, the greater is the response of the membrane. The electrodes exhibit good tolerance toward sugars, amino acids, and urea as the presence of these species up to 4- or even 5-fold excess did not affect significantly the potential reading. This has indicated that the presence of these species can be tolerated into a high extent. Such a high tolerance can then be attributed to the differences in polarity and lipophilic nature of their molecules relative to those of the drugs under investigation. Though all *Olan*, *Oxaz* and *Lora* mutually interfere, these three similar substances so far do not exist in one pharmaceutical preparation.

3.3. Effect of pH and of the Type of Buffer

The influence of pH on the response of the proposed drug-selective electrode was checked by measuring the potential displayed by 1×10^{-3} and 1×10^{-4} M drug test solutions over the range of pH 2-11. Small volumes of dilute sodium hydroxide and / or HCl were used for adjustment. It is apparent from the potential-pH profiles that the response of all three sensors was fairly constant in acetate buffer solutions of pH 3.5-6.0. The potential did not vary over this range by more than ± 3 mV and the electrode could be applied for the determination of investigated drugs. A considerable decrease of the potential above pH 6 was observed, which was probably due to the decreased concentration of the protonated form of the drug, whereas — at higher pH — a deviation in the potentials reading might occur due to the penetration of the hydroxonium ion into the membrane layer. On the other hand, two types of buffers, i.e., acetate and phosphate media were investigated on *Olanzapine*-ISE in order to ensure that the electrode would be functioning [23]. The effect of the two buffers was not critical and no marked difference was observed as shown in Fig. 7 and Table 5. However, acetate buffer of pH 4.5 was ascertained to be more suitable because of the fact that the sensor potentials were almost constant and stable within ± 3 mV.

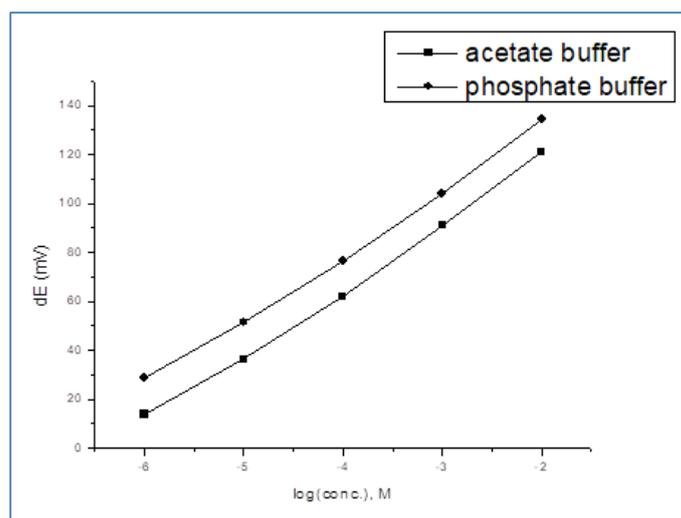


Figure 7 Calibration curves of *Olan*-electrode obtained in acetate and phosphate buffers

3.4. Effect of the Plasticizer Used

In this work, poly-vinyl chloride (PVC) membranes plasticized with DBS or NPOE and containing the lipophilic salt, *Olan-Re*, *Oxaz-Re* or *Lora-Re*, were prepared. Potentiometric responses of the electrodes based on neutral ionophores and modified with the investigated drugs were greatly influenced by the polarity of the membrane medium, which was in turn defined by the dielectric constants of the incorporated drug moiety and plasticizer (solvent mediator) used. Regarding the latter, both dielectric constants were $D_{i(\text{DBS})} = 4.01$ and $D_{i(\text{NPOE})} = 23.6$, indicating a high sensitivity and nearly Nernstian slope for the ISEs with NPOE-plasticized membrane, whereas the ISEs with DBS-plasticized membranes showed anomalous response to the corresponding drug.

3.5. Composition of the Membrane

In plastic membranes of ISEs studied, the amount of the ion-pair exchanger should be sufficient to obtain reasonable ionic exchange at the gel layer-test solution interface, which is responsible for the membrane potential and its changes. Also, the amount of the plasticizer should be the extent that produces a membrane of good physical properties and at the same time plays efficiently its role as a solvent mediator for the ion-exchanger.

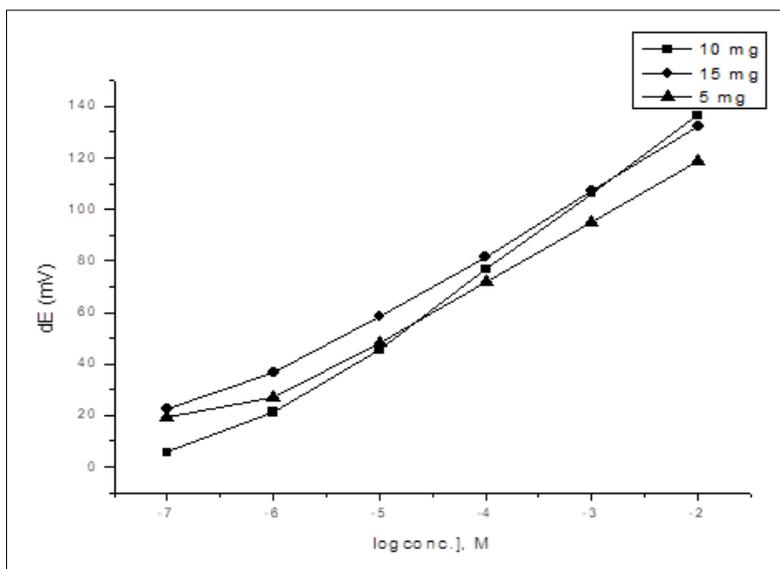


Figure 8 Effect of the composition of ionophore on the response of *Olan*-sensors

The composition of each of these membranes was varied in such a way in order to reach the optimum composition exhibiting the best performance characteristics, (calibration-curve slope, rectilinear concentration range, and the response reproducibility). Further, it comprised the different compositions of ionophore cooperating in the membrane and the slope (mV/decade) of the obtained calibration graphs for *Olan*-sensor plasticized with NPOE. The membrane was prepared three times and the relative standard deviation (R.S.D.) calculated for the slope values was always fairly low, showing on good reproducibility of the preparation process and comparable slope values of the calibration graphs obtained with ISEs differing in composition. As shown in Fig. 5, it is evident that the change of composition of ionophore has significantly affected the response of the electrode. The proper results are gathered in Table 4, showing that the optimal composition of ionophore is 10 mg, offering an ISE with the best performance.

Table 4 Effect of ionophore on the response of *Olan*-ISE with NPOE as plasticizer

Composition [mg]	5	10	15
Parameters evaluated			
Slope, (mV/decade)	20.75	26.91	22.38
Intercept	156.67	186.25	173.66
R.S.D. [% , ±]	5.1	3.83	4.6
Limit of detection [mol L ⁻¹]	6.2×10^{-7}	3.1×10^{-7}	4.6×10^{-7}
Correlation coeff. (r ²)	0.9839	0.9988	0.9923

3.6. Regeneration of the Ion-Selective Electrodes Prepared

From the above discussion, it is clear that soaking of the electrodes in their respective drug solutions for a long time has a negative effect on the response of the membrane towards the ions investigated. The same effect appears after working with the electrode for a long time. The regeneration of such "exhausted" electrodes was tried by simply re-loading of

the ion-exchangers of the *Olan*-RE, *Oxa*-RE, and *Lora*-RE type, in the external gel layer of the membrane [24]. This was accomplished by soaking the drained electrodes for 24 hours in a solution of 0.01 M A-RE, followed by saturation in a solution of the respective drug. As found, the soaking in 0.01 M solution of each drug for ca. three hours was sufficient for regeneration and seen from the slopes of the renewed electrodes, increasing from 21.32, 45.70, and 43.33 up to 24.51, 52.16 and 50.01 mV/decade for *Olan*-, *Oxaz*-, and *Lora*-ISEs, respectively. Generally, it has been found that the life time of regenerated electrodes is limited up to few hours (60-180 min.). This was due to the ease of leaching of lipophilic salts from the gel layer at the electrode surface compared to a situation, when they had been firmly attached to the PVC network through the solvent mediator with strong binding forces.

3.7. Choice of the Method for Quantitative Analysis and Potentiometric Determination of the Drugs of Interest Using BZD/RE-modified PVC-Membrane ISEs.

In fact, three principal methods could be considered as potentially applicable to the quantitative analysis with the three *Drug*/RE-ISEs developed during these studies. Such a choice comprised: (i) direct calibration measurements, construction of the corresponding plot and the calculation of the concentration from the Nernst equation; (ii) potentiometric titration involving the use of counter ions as the titrant which seemed more accurate, depending essentially on the use of the ISE as end-point detector, but more time consuming (in an estimate, at least 20-30 min. for the average analysis); (iii) the standard addition(s) method, which is often most reliable, due to the most effective suppression of matrix effects, and therefore quite frequently applied in practical determinations with ISEs.

Direct Potentiometry: The individual drugs contained in commercial pharmaceutical preparations had represented the first part of practical samples that were analyzed and their concentrations determined with the respective ISE when using the standard addition method.

The results obtained are listed in Table 5. By applying the least-squares method for five replicate determination (i.e., n = 5) at a 95% confidence level, the corresponding regression equations can be written as follows:

$$Y = 0.984(\pm 0.0037) X + 0.0133(\pm 0.0718) \quad \dots \text{ for Olanzapine} \quad (2a)$$

$$Y = 1.003(\pm 0.0060) X - 0.2699(\pm 0.1076) \quad \dots \text{ for Oxazepam} \quad (2b)$$

$$Y = 0.995(\pm 0.3101) X - 0.2138(\pm 0.0137) \quad \dots \text{ for Lorazepam} \quad (2c)$$

When all three regression plots have exhibited nearly ideal correlation (with $r^2 = 0.9999$) and where "X" is the average reference assay and "Y" is the average found by the proposed electrode / method. Also, the RSDs obtained were highly satisfactory, ranging from ± 0.8 to ± 2.1 %, and the same can be stated about the recovery rates varying from 97.93 to 99.27 %, with the variation coefficients within an interval of 0.81-1.66. Finally, there is also a fine agreement with the results of the official reference method [25].

The results show that the proposed electrodes can be used to determine *Olanzapine*, *Oxazepam* and *Lorazepam* in pure samples or pharmaceutical preparations with both high accuracy and high recovery; all without time consuming pre-treatment procedures required, e.g., to minimize the undesirable matrix effects. Also, the target drugs were successfully determined in spiked urine samples with the mean recoveries within 97.67-99.31 %, with the RSDs in an interval of 0.16-0.53 %, respectively

Table 5 Quantification of the content of *Olanzapine*, *Oxazepam*, and *Lorazepam* in pharmaceutical preparations using the respective ISE with NPOE as plasticizer

Taken, [$\mu\text{g/ml}$]	Method proposed			Reference method*		
	Recovery [%]	R.S.D C.V.	t-value	Recovery [%]	R.S.D. [% , \pm]	F
Olanzapine, Zyprexa tablet 10 mg						
12.50	98.40	0.20 1.62	0.6	99.11	0.56	1.14
18.75	98.66	0.15	0.85	98.32	0.41	0.13

		0.81				
25.00	98.42	0.41	2.34	100.11	0.37	1.22
		1.66				
Oxazepam , Serenid-D (Weyth) tablet, 15 mg						
11.46	98.22	0.43	1.10	100.02	0.33	1.69
		0.83				
17.20	98.58	0.47	1.18	97.12	0.40	1.38
		1.27				
22.93	99.27	0.36	0.86	98.65	0.25	2.07
		1.51				
Lorazepam , Lorazem tablet, 2 mg						
12.85	97.93	0.26	1.80	99.54	0.22	1.39
		1.20				
19.30	98.44	0.66	1.18	100.21	0.54	1.49
		1.42				
29.70	98.83	0.44	0.76	98.35	0.34	1.67
		1.49				

The recovery and the RSDs characterizing the method tested are summarized in Table 7, surveying also other useful details. These results have shown that all the electrodes can detect the drugs in the spiked urine samples with high accuracy and precision, as well as with a high recovery; again, without any additional pre-treatment procedures.

Table 6 Potentiometric determination of *Olanzapine*, *Oxazepam*, and *Lorazepam* in urine samples using the respective ISE with NPOE as plasticizer

Taken [µg/ml]	Proposed method			Reference method*		
	Recovery [%]	R.S.D. [% , ±]	t-value	Recovery [%]	R.S.D. [% , ±]	F
Olanzapine						
12.50	97.67	0.50	0.27	98.11	0.56	0.79
18.75	97.78	0.41	0.53	98.32	1.03	0.15
25.00	98.23	0.47	2.36	100.21	0.25	3.52
Oxazepam						
11.46	97.68	0.26	1.72	99.42	0.33	0.63
17.20	98.12	0.32	0.22	98.00	0.23	1.93
22.93	99.31	0.49	0.68	98.65	0.36	1.85
Lorazepam						
12.85	98.24	0.16	1.37	99.54	0.22	0.53
19.30	99.12	0.40	1.15	100.21	0.34	1.38
29.70	98.00	0.53	0.41	98.35	0.34	2.42

The results outline that the proposed electrodes can be used to determine *Olanzapine*, *Oxazepam* and *Lorazepam* in pure samples or pharmaceutical preparations with both high accuracy and high recovery; all without time-consuming pre-treatment procedures required e.g. to minimize the undesirable matrix effects. Also, the target drugs were successfully determined in spiked urine samples with the mean recovery values, ranging from 97.67 to 99.31 % with the RSDs in an interval of 0.16-0.53 %, respectively. The recovery and the RSDs characterising the method tested are summarised in Table VII, surveying also other useful details. These results have shown that all the electrodes can detect the drugs in the spiked urine samples with high accuracy and precision, as well as with a high recovery; again, without any additional pre-treatment procedures.

Potentiometric Titrations: Though the determination by potentiometric titrations may be time consuming, it offers the advantage of high accuracy and precision, when the end-point can easily be determined via the sharp potential break. In addition, partially exhausted electrode can also be employed because the actual / exact potential value during titration and at its end-point is of secondary interest [26]. Representative titration curves for the determination of all three drugs using the respective electrodes are shown in Figs. 9, 10, and 11. Notable is the fact that as the concentration of the drug had increased, the inflection of the break-point became sharper than those for lower concentrations of the drugs. Then, these electrodes could be successfully used to indicate properly the potentiometric titrations of the drugs investigated.

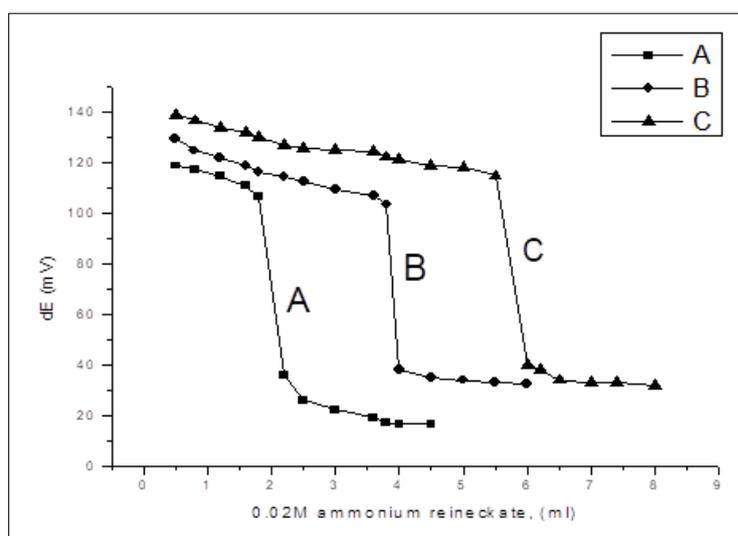


Figure 9 Typical potentiometric titration curves of (A) 2.0 mL, (B) 4.0 mL, and (C) 6 mL 0.01 M *Olanzapine* with a standard solution of 0.01 M ammonium reineckate (A-RE) as the titrant and using *Olan*-ISE as indicator electrode

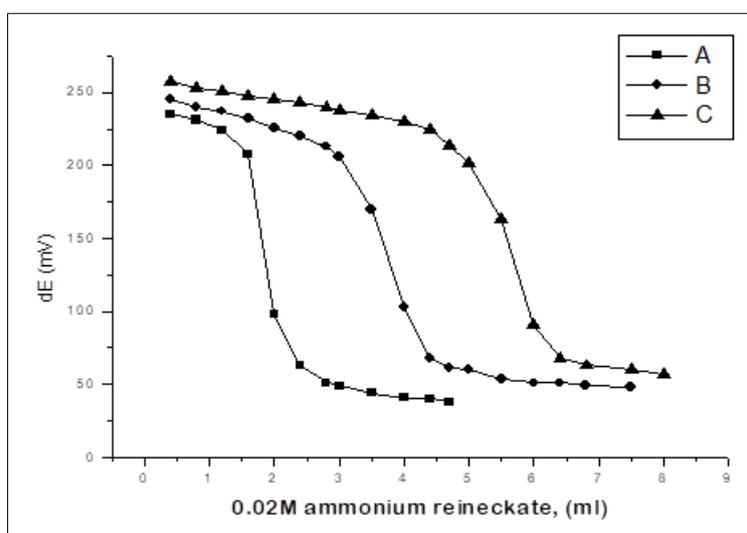


Figure 10 Typical potentiometric titration curves of (A) 2.0 mL, (B) 4.0 mL, and (C) 6 mL 0.01 M *Oxazepam* with a standard solution of 0.01 M A-RE using *Oxaz*-ISE

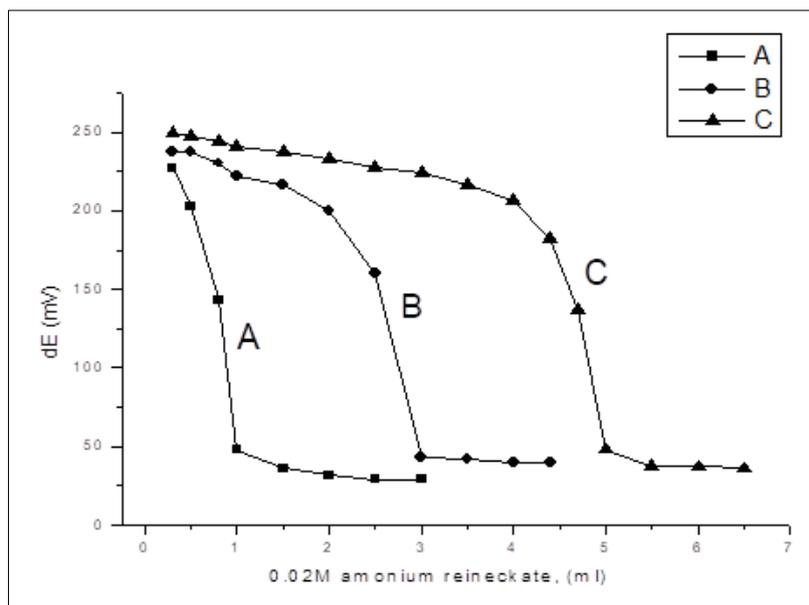


Figure 11 Typical potentiometric titration curves of (A) 1.0 mL, (B) 3.0 mL and (C) 5 mL 0.01 M *Lorazepam* with a standard solution of 0.01 M A-RE using *Lora*-ISE.

3.8. Statistical Evaluation of Both Methods for Quantitative Analysis

The results of conventional F- and t-tests are shown in Table 7. The obtained values were substantially lower than the theoretical (tabulated) values; i.e., the methods applied do not exhibit significant differences compared to those obtained by the official (reference) method.

Table 7 Statistical treatment of the data obtained from the determination of the three drugs at the electrodes proposed in comparison with the reference method employing HPLC

Parameters Evaluated	Reference method (based on HPLC)	Olanzapine electrode	Oxazepam electrode	Lorazepam electrode
Working range [M]	0.5-18 µg/ml *	10 ⁻⁷ -10 ⁻²	10 ⁻⁶ -10 ⁻²	10 ⁻⁶ -10 ⁻²
	12-40 µg/ml **			
	20-130 µg/ml **			
LOD *** [mol L ⁻¹]	0.12 µg/ml *	3.2 × 10 ⁻⁷	4.6 × 10 ⁻⁶	3.7 × 10 ⁻⁶
	3.5 µg/ml **			
	5.0 µg/ml **			
Accuracy [%]	98.89±1.3 *	98.40-98.66	98.22-99.27	97.93-98.83
	98.56±1.1 **			
	98.41±1.3 **			

* Reference method (used) for *Olanzapine* (see Ref. [25]); ** Reference method for *Oxazepam* and *Lorazepam* [25]; *** Limit of detection; average of three determinations

To check the accuracy, the new method (with the electrodes tested) was compared with standard method in an effort to find if there is any significant difference which could reveal the extent into which potential deviation(s) may affect the applicability of the new method(s) compared to the already existing method(s). As found out, the method examined did not exhibit any significant differences, as confirmed by the data in the last Table.

4. Conclusions

The new ISEs based on the functioning of the BZD / RE ion-associate in the PVC-membrane have been shown to be applicable in potentiometric analysis of three pharmaceuticals of the benzodiazepine family; specifically, they allow one to determine as low as 1×10^{-7} M *Olanzapine* and 1×10^{-6} M *Oxazepam* or *Lorazepam*. The electrodes proposed offer the advantages of low cost, ease of fabrication, high stability, fast response over a wide concentration of the analyte(s) and almost independent of pH, as well as adequate selectivity in the presence of related species.

The whole procedure has also been found sufficiently reliable for quantitative analysis, when combined either with calibration curve method or with standard addition(s). Last but not least, the drugs of interest can be determined either in pure (powdered) form or in the commercial dosage (tablet) forms without any special pre-treatment and the employment of all three ISEs developed seems to be feasible also in analyses of turbid or intensively coloured sample solutions. It can be concluded that the procedure described in the previous sections is a valuable contribution to the palette of similar methods already developed for analysis of pharmaceuticals with familiar structures, showing also that potentiometric indication may offer a possible alternative to more common voltammetric measurements.

Compliance with ethical standards

Disclosure of conflict of interest

No conflict of interest to be disclosed.

References

- [1] Atropisomers of 1,4-benzodiazepines. Synthesis and resolution of a diazepam-related 1,4-benzodiazepine .Norman W. Gilman, Perry Rosen, James V. Earley, Charles Cook, and Louis J. Todaro J. Am. Chem. Soc. **112**, 3969 (1990)
- [2] Double-blind, randomized comparison of olanzapine versus fluphenazine in the long-term treatment of schizophrenia. Dossenbach M., Folnegovic V. S., Hotujac L., Uglesic B., Tollefson G., Grundy, S., Friedel P., Jakovljevic M.: Prog. Neuropsychopharmacol. Biol. Psychiatry **28**, 311 (2004)
- [3] High-dose glycine added to olanzapine and risperidone for the treatment of schizophrenia Uriel Heresco-Levy 1, Marina Ermilov, Pesah Lichtenberg, Gali Bar, Daniel C Javitt : Biol. Psychiatry **55**, 165 (2004).
- [4] Effectiveness and medication acceptance of olanzapine disintegrating tablets compared to standard olanzapine tablets in acutely treated psychiatric patients Sinclair D. A., Skaer T. L., Robinson L. M., Dickson W. M., Markowitz J. S., De Vane C. L.: Eur. Neuropsychopharmacol. **13**, S281 (2003).
- [5] Clarkes J.: *Analysis of Drugs and Poisons*. Wiley, N.Y. (2005)
- [6] Moller H. J.: J. Clin. Psychopharmacol. **19**, 2S (1999).
- [7] Lader M. H.: Eur. Neuropsychopharmacol. **9**, 399 (1999).
- [8] Ashton H.: *Textbook of Drugs*. Elsevier, Amsterdam (1994).
- [9] Argyropoulos SV, Nutt DJ,. The use of benzodiazepines in anxiety and other disorders,.: Eur. Neuropsychopharmacol. **9**, 407 (1999)
- [10] Treatment of Men With Major Depression: A Comparison of Sequential Cohorts Treated With Either Cognitive-Behavioral Therapy or Newer Generation Antidepressants. Verwey P., Eling H., Wientjes F., Zitman G.: J. Clin. Psychiatry **61**, 465 (2000).
- [11] Epidémiologie de la consommation des anxiolytiques et des hypnotiques en France et dans le monde Pelissolo G.: Encephale **22**, 187 (1996)
- [12] Pharmacokinetic and pharmacodynamic analysis of sedative and amnesic effects of lorazepam in healthy volunteers . Blin N., Simon E., Jouve M., Habib D., Gayraud A., Durand, B., Bruguerolle, P. Pisano: Clin. Neuropharmacol. **24**, 71 (2001).
- [13] A double-blind, randomized comparison of i.v. lorazepam versus midazolam for sedation of ICU patients via a pharmacologic model. Barr J., Zomorodi K., Bertaccini E. J., Shafer S. L., Geller, E.: Anesthesiology **95**, 286 (2001).

- [14] USE OF LORAZEPAM AS A PREMEDICANT FOR CAESAREAN SECTION: An evaluation of its effects on the mother and the neonate Houghton J.: *Brain J. Anaesth.* **55**, 767 (1983).
- [15] Clinical Pharmacokinetics of Sedatives in Neonates. Jacaz A., Burtin P.: *Clin. Pharmacokinet.* **31**, 423 (1996)
- [16] U.S. Norm, "USP-NF27", United States Pharmacopoeia Convention, Rockville (2008). Hassan S.M., Abu Sekkina M.M., El Ries M. A.: *J. Pharm. Biomed. Anal* **30**, 175 (2003).
- [17] Authors' coll.: *British Pharmacopoeia*. London: HMSO Publication (2007).
- [18] Direct injection HPLC method for the determination of selected benzodiazepines in plasma using a Hisep column .Pistos C., Stewart James T.: *J. Pharm. Biomed. Anal.* **33**, 1135 (2003)
- [19] U.S. Norm, "USP-NF27", United States Pharmacopoeia Convention, Rockville (2008) Sokalski T., Ceresa A., Zwickl T., Pretsch E.: *J. Am. Chem. Soc.* **119**, 11347 (1997).
- [20] Skoog D. A., Holler E. J., Nieman T. A.: *Principles of Instrumental Analysis*, 5th Ed.; Harcourt & Brace, Orlando (1998).
- [21] Phenothiazine drug poly(vinyl chloride) matrix membrane electrodes and their use in pharmaceutical analysis Cosofret V. V., Buck R. P.: *Analyst* **109**, 1321 (1984).
- [22] Determination of selectivity coefficients of ion-selective electrodes by a matched-potential method. Gadzekpo V. P. Y., Christian G. D., *Anal. Chem. Acta* **164**, 279 (1984).
- [23] A Polymer Membrane Sensor for Silver. Bates M. M., Cardwell T. J., Cattrall R. W., Deady L. W., Gregorio C. G.: *Talanta* **42**, 999 (1995).
- [24] Performance characteristics and regeneration of a tetraphenylboron(III) selective electrode. Shoukry A.F., Badway S.S., Issa Y.M.: *Anal. Chem.* **59**, 1078 (1987).
- [25] Fritz F. S., Schenk G. H.: *Quantitative Analytical Chemistry*, 4th Ed.; Allyn & Bacon, London (1975). Pistos C., Stewart J.T.: *J. Pharm. Biomed. Anal.* **33**, 1135 (2003); and *United States Pharmacopoeia*, USP-NF27; U. S. Pharm. Convention Inc., Rockville Md., (2008).
- [26] Fritz F.S., Schenk G.H.: *Quantitative Analytical Chemistry*, 4th Ed.; Allyn Bacon, London, 1975.
- [27] El Maali N.: Voltammetric analysis of drugs, *Bioelectrochem.* **64**, 99 (2004).