

The interaction between neuroendocrine and immune systems in infants and children during under nutrition

Ashraf Soliman ^{1,*}, Nada Alaaraj ¹, Doaa K Yassin ² and Alan D. Rogol ³

¹ *Pediatric Diabetes and Endocrinology, Hamad general hospital, Qatar.*

² *Pediatric Endocrine and Diabetes Unit, Faculty of Medicine, University of Alexandria, Alexandria, Egypt.*

³ *Department of Pediatrics, Professor Emeritus, Division of Diabetes and Endocrinology, University of Virginia, Virginia, USA.*

World Journal of Advanced Research and Reviews, 2023, 18(03), 413–424

Publication history: Received on 01 May 2023; revised on 08 June 2023; accepted on 12 June 2023

Article DOI: <https://doi.org/10.30574/wjarr.2023.18.3.1104>

Abstract

Malnutrition remains an international concern because it represents a major cause of childhood morbidity and mortality. In infants and children malnutrition significant metabolic, hormonal, and immune changes occur with short- and long-term effects and outcomes. Thus, studies which contribute to the prevention, amelioration, or rehabilitation of the detrimental effect of malnutrition are necessary. Investigations on endocrine, metabolic, and immune changes related to severe malnutrition started a long time ago. However, recently there has been little work on these important changes, their mechanism/s of action and interactions, and their effect on different systems. The neuroendocrine system is the initial response to stress, so it is reasonable to assume that this system also plays a key role in the pathophysiological changes during nutritional deprivation. However, many of the newly identified functions/changes are better explained by the action of conventional neurotransmitters (e.g., glutamate and GABA) that constitute a neuronal circuit. In addition, the mechanistic Target of Rapamycin (mTOR) is an evolutionarily conserved serine/threonine kinase has emerged as a sensor for nutrients that has a central joint in cellular metabolism, cell growth, and differentiation. Its functions during malnutrition in the central nervous system, endocrine system, and different end organs and tissues (liver, pancreas, muscle, and adipose tissue) need elucidation. This article presents a brief review of neuroendocrine changes during malnutrition and their effects on the modulation of metabolism, growth, and immune functions. The pros and cons of these endocrine changes are discussed as well as their reversibility on nutritional rehabilitation.

Keywords: Neuroendocrine; Hormones; Immune system; Growth; Infection; Inflammation; Enteropathy

1. Introduction

About 8 million of the world's children under the age of 5 years die every year; undernutrition is the underlying factor in 35% of these cases (1–3). The consequences of chronic malnutrition in children are both immediate and long-term and include increased morbidity and mortality, poor child growth and development, impaired learning capacity, and increased risk of infections. In addition, they have an increased risk of developing non-communicable diseases including obesity, diabetes, hypertension, and dyslipidemia. During their adulthood, they have lowered working capacity and unfavorable maternal reproductive outcomes. (3-7) However, obesity, as a form of malnutrition, is not a part of this review.

The neuroendocrine system including the hypothalamus, and pituitary gland, and their network connections represent a major regulatory system. The hypothalamic hormones control the secretion of the anterior pituitary hormones which are necessary for adaptation to internal and external stresses, growth, reproduction, and metabolism. Body homeostasis

* Corresponding author: Ashraf Soliman

including linear growth and weight gain is predominantly controlled by hormones secreted by endocrine organs. In addition, the neuroendocrine system also cooperates with the immune system in regulating body responses to various forms of stress and in modulating responses to stimuli provoking immunologic responses. (8,9)

Conventionally, neurohormones released by the hypothalamus and the pituitary gland have received much consideration owing to the distinctive roles of the end hormones released by their target peripheral organs (e.g., glucocorticoids, growth hormone, thyroxine). (10)

Recent advances have revealed several important metabolic functions of hypothalamic neurohormone-expressing cells, many of which are not readily explained by the action of the corresponding classical downstream hormones. These newly identified functions are better explained by the action of conventional neurotransmitters (e.g., glutamate and GABA) that constitute a neuronal circuit. For example, leptin (secreted by adipose cells) plays an important role in regulating energy balance largely through acting on γ -aminobutyric acid (GABA)ergic neurons in the brain. Ghrelin is an orexigenic hormone primarily produced in the stomach and increases appetite as well as growth hormone (GH) release. Ghrelin antagonizes leptin by reducing the firing of proopiomelanocortin (POMC) neurons by raising the frequency of spontaneous synaptic γ -aminobutyric acid (GABA) release onto them. (11-12)

In addition, the mechanistic Target of Rapamycin (mTOR) is an evolutionarily conserved serine/threonine kinase which is a member of the PI3K related kinase (PIKK) family, has emerged as a sensor for nutrients that has a central joint in cellular metabolism, cell growth, and differentiation, in the central nervous system as well in different tissues (liver, pancreas, muscle, and adipose tissue). (13)

The malnutrition-related immune suppression increases susceptibility to infection and is characterized by recurrent infections and chronic inflammation. Both infection and inflammation aggressively contribute to malnutrition which causes and perpetuates a vicious cycle. Death rates from diarrhea, pneumonia, measles, and malaria are increased considerably in undernourished children. (14) Malnutrition causes immunosuppression through a variety of peripheral and central mechanisms, including the involvement of the central nervous system and endocrine system.

Investigating the possible relation/s between changes in the neuroendocrine system and immune system represents an important step to understand the pathophysiologic mechanisms that contributes to lowered systemic and local immunity during chronic malnutrition. (15,16)

Classical studies analyzed the levels of selected hormones and metabolites in undernourished infants and young children and healthy adults subjected to prolonged fasting. Yet link between these hormonal changes and the pathogenesis of malnutrition remains inadequately recognized. In this review, we studied the changes of the neuroendocrine system reported by different investigators and discussed their possible effects (pros and cons) on body homeostasis (metabolic, growth and immune functions) in infants and children during severe malnutrition as well as the effect of nutritional rehabilitation on these changes. (17-21)

2. Discussion

2.1. General perspectives

The neuroendocrine system is the initial response to stress, so it is reasonable to assume that this system also plays a key role in the pathophysiological changes during nutritional deprivation. In addition to their metabolic effects, recent research suggested and proposed an important role of hormonal changes in the control of growth, musculoskeletal system, immunity, inflammation, and response to infection. (9) Activation of the HPA axis provides a slower, sustained, and amplified physiological response to stress including starvation. Stressor perception is communicated to neurons in the PVN which in turn release corticotropin-releasing hormone (CRH) and other releasing factors, from in the median eminence. CRH passes through the hypothalamic-pituitary portal circulation to the pituitary and stimulates the release of adrenocorticotrophic hormone (ACTH) from corticotropes into the systemic circulation. Subsequently, ACTH stimulates the adrenal cortex to produce and secrete cortisol. Glucocorticoids act on their receptors throughout the brain and body to exert numerous effects, including mobilization of stored energy. Glucocorticoids mobilize fuel from the liver via increased gluconeogenesis and from white adipose tissue by increased lipolysis. The net effect is to increase the availability of fuel, facilitating a physiological response to the (starvation) threat. (22-24)

The central nervous system and the autonomic system combine and integrate multiple peripheral signals to control appetite. Peripheral hormonal messages indicative of long-term energy whole-body status are produced by adipose tissue including leptin, and adiponectin. On the other hand, acute orexigenic (+) ghrelin signal (produced in the gut) and

anorexigenic (–) signals such as the gut hormones peptide YY (PYY), glucagon-like peptide-1 (GLP-1), and cholecystikinin (CCK), and the pancreatic hormones [insulin, glucagon, amylin, and pancreatic polypeptide (PP)] indicate long-term energy status. Proopiomelanocortin (POMC) neurons mainly located in the arcuate nucleus of the hypothalamus and the nucleus tractus solitarius of the brainstem are the site of this control. POMC neurons are inhibited by energy deficits. During fasting (starvation), for instance, when both glucose and insulin levels drop, certain POMC neuronal subsets could either be activated or inhibited, working towards the same goal of increasing plasma blood glucose or reducing glucose uptake into peripheral organs, to maintain constant systemic glucose levels. (25-26)

The functions of the mechanistic mTOR system during malnutrition. In addition to the central mechanisms regulating endocrine, growth, and immunological changes during nutritional deprivation stress, control through the Target of Rapamycin (mTOR) represents a major cellular control mechanism. The mechanistic mTOR is an evolutionarily conserved serine/threonine kinase that is a member of the PI3 K-related kinase (PIKK) family. mTOR emerged as a central joint in cellular metabolism, cell growth, and differentiation. It senses the nutrients, energy, insulin, growth factors (GH and IGF1), and environmental signals and transmits signals to downstream targets to downregulate nutrient and energy use and helps cells survive periods of starvation and intermittent fasting. During malnutrition shortage of nutrients (amino acids, glucose) and lower insulin and IGF1 decrease, the mTOR activity which decreases the mTOR mediated cellular protein synthesis and lipogenesis and epiphyseal cartilage growth. Notably, mTOR has been also implicated in the regulation of both the innate and adaptive immune responses. (27-29)

In the liver, this inhibition of mTOR is required for inhibiting lipogenesis and activation of ketogenesis and it regulates the hepatic glucose output and peripheral lipid metabolism. In skeletal muscle, mTORC1 activity exerts a significant effect on muscle mass by affecting protein synthesis and degradation (autophagy process). mTOR inhibition increases the rate of protein degradation and decreases protein synthesis. (Muscle wasting) and decreases skeletal muscle insulin sensitivity. In addition, glucocorticoids may regulate mTOR by modulating the level of both BCAT2 and myostatin to regulate catabolism in skeletal muscle. (30-32)

In the brain, within the hypothalamus, mTOR functions as a cellular signaling hub that integrates internal and external cues (which is associated with changes in energy status) to control the central or peripheral tissue functions. mTOR plays a critical role in the regulation of food intake and body weight. mTOR combines and coordinates signals from various "energy controlling" hormones such as leptin, insulin, IGF-I, and ghrelin. This regulatory mechanism occurs in the arcuate nucleus (ARC) of the hypothalamus. Peripheral hormones such as ghrelin and leptin act on 2 types of neurons that can analyze the metabolic status of the periphery: 1. The orectic neurons [neuropeptide Y (NPY)/agouti-related peptide (AgRP) containing neurons and 2. The anorectic neurons [POMC and cocaine- and amphetamine-regulated transcript (CART)-containing neurons]. (32-34) On the other hand, inhibition of mTOR negatively affects immunity because mTOR plays a central role in the differentiation of T-cell subsets, and controls aspects of B-cell and antigen-presenting cell (APC) development. (32-36)

There are bi-directional circuits linking CNS and immune systems. The CNS can connect with the immune system to regulate its activity, through the autonomic nervous system, the catecholaminergic pathway, or the neuropeptides, and through controlling anterior pituitary hormone release. In this perspective, GH and IGF1, corticosteroids, insulin, and leptin can modulate the immune system by different mechanisms. (9)

3. Hormonal changes and their effect on metabolism, growth, and immunity during malnutrition: (figure 2)

3.1. Adipocyte Hormones

3.1.1. Leptin

Leptin secretion and its level in blood are low in all forms of malnutrition (mild, moderate, and severe) due to lower or depleted fat mass. (36-38) The leptin level serves as a measure of energy reserves (adipose tissue) and instructs the central nervous system to manipulate food consumption and energy spending accordingly. In response to fasting and starvation, leptin levels fall rapidly before and out of proportion to any changes in fat mass. Low leptin levels have been documented in all studies of children with malnutrition despite the lack of standardization of the timing of the collection of samples. (39) The fall in serum leptin concentration leads to neurohumoral and behavioral changes, trying to preserve energy reserves for vital functions. Emerging evidence suggests that starvation hypo-leptinemia increase the activity of the hypothalamic-pituitary-adrenal axis, promoting white adipose tissue (WAT) lipolysis, increasing hepatic acetyl-CoA concentrations, and maintaining euglycemia. In addition, leptin is also responsible for facilitating the shift from a dependence upon glucose metabolism (absorption and glycogenolysis) to fat metabolism (lipolysis increasing

gluconeogenesis). This function sustains the supply of energy substrates to the brain, heart, and other vital organs. In this way, a leptin-mediated glucose-fatty acid cycle appears to maintain glycemia and permit survival in starvation. (37-40)

Leptin promotes IFN- γ secretion by memory T cells, inhibits Th2 responses, and induces activation markers (CD69, CD25, and CD71). In addition to inducing lymphopoiesis, leptin seems to deliver survival signals to T cells (94) In innate immunity, leptin enhances the activity of neutrophils by the release of oxygen free radicals. It stimulates the migration of immune cells at the sites of inflammation through increasing intercellular adhesion molecule-1 (ICAM-1). Moreover, leptin activates the monocytes and dendrite cells (DCs) that in turn leads to the production of pro-inflammatory cytokines such as TNF- α , and IL-6 along with IL-12, a key cytokine that facilitates the shifting of T-cells toward the Th1 phenotype. Leptin also promotes DCs survival by triggering the activation of nuclear factor-kappa B (NF-kappa B) (41-44)

These important functions explain how leptin deficiency negatively influences cellular immunity and predisposes to infection. Leptin deficiency in both mice and humans results in severe immune defects characterized by a decrease in total lymphocytes, CD4+ helper T cell number, increased thymocyte apoptosis, and a shift from the Th1 toward Th2 phenotype. These changes increase susceptibility to intracellular infections. The systemic leptin deficiency in malnutrition correlates with several bacterial, viral, and parasitic infections due to defective cytokine production. These include tuberculosis, pneumonia, sepsis, colitis, viral infection, leishmaniasis, trypanosomiasis, amoebiasis, and malaria. (44) In 77 edematous and non-edematous undernourished children a diminished leptin level was the most reliable biochemical predictor of mortality. (45,47)

3.1.2. Adiponectin

Bartz et al reported decreased Adiponectin in their undernourished children which increased significantly on nutritional rehabilitation. (46) Adiponectin is one of the key adipocyte-derived hormones that regulate systemic or tissue lipid and glucose metabolism. Contrary to the action of other adipocyte-derived hormones, adiponectin improves insulin sensitivity and enhances lipid and glucose metabolism. It increases fatty acid oxidation in the liver and muscle. It promotes insulin-sensitizing and fat-burning which can be useful during malnutrition. Adiponectin plays a role in insulin sensitivity by impacting insulin sensitivity in skeletal muscle and liver and increasing insulin release from beta cells. Low adiponectin (secreted by adipocytes) may lead to impaired insulin secretion and decreased insulin sensitivity in these children. (48-51) The anti-inflammatory properties of Adiponectin are due to its suppression of M1 macrophage activation and supporting M2 macrophage proliferation. It decreases inflammation, apoptosis, and oxidative injury in muscle, heart, and brain. Its decrease during malnutrition may encourage the inflammation process and increase oxidative tissue injury. (52)

3.2. Gut hormones

The gastrointestinal (GI) tract is the body's largest endocrine organ producing hormones that have important sensing and signaling roles in the regulation of energy homeostasis. (53) Ghrelin is a multifaceted gut hormone with many functions including stimulating food intake, fat deposition, and growth hormone release. Ghrelin regulates glucose hemostasis by inhibiting insulin secretion and regulating gluconeogenesis/glycogenolysis. Ghrelin shares in maintaining glucose homeostasis during starvation through increasing glucose synthesis and release by activating gluconeogenesis and/or increasing growth hormone. Bartz et al reported hyper-ghrelinemia in undernourished children. (46) The high Ghrelin stimulates GH secretion which promotes lipolysis and maintains blood glucose through stimulating hepatic gluconeogenesis. The lipolytic and gluconeogenic effects of GH are independent of IGF-I. (53-56) GLP-1 is secreted in the small intestine in response to nutrients. It promotes glucose-dependent insulin secretion, decelerates gastric emptying, and reduces food intake.

On the other hand, intraluminal nutrients, particularly fats stimulate the secretion of PYY by enteroendocrine cells. PYY inhibits gastric emptying and induces satiety. Levels of GLP-1 and PYY at baseline were found to be considerably higher in undernourished infants compared to normal infants and children and declined sharply during outpatient RUTF treatment. The significant increase of these GI hormones during malnutrition helps to delay gastric emptying and give longer time for nutrient absorption. Low levels of leptin and adiponectin and high levels of PYY were correlated with mortality in these children. (57)

Many undernourished children living in low-income countries (LICs) suffer from environmental enteric dysfunction (EED). EED refers to a subclinical enteropathy characterized by mucosal inflammation and villus blunting mediated by T cell activation. Exposure to intestinal pathogens and intestinal dysbiosis (an imbalance of the microorganisms within the intestine), as a consequence of poor sanitation and possibly specific micronutrient deficiencies (e.g, zinc, vitamin D,

vitamin A, and protein), lead to intestinal inflammation and disruption of intestinal barrier function. The damaged barrier function permits the translocation of pathogens (bacteria and bacterial products from the intestine to the mesenteric and systemic circulation. This exposure activates innate immune cells in the mesenteric lymph nodes, liver, and systemic circulation to generate proinflammatory cytokines. (58,59)

Zonulin is a hormone secreted mainly from the liver, but also from enterocytes, adipose tissue, brain, heart, immune cells, lungs, kidney, and skin. It is a master modulator of the intercellular tight junctions, important in antigen trafficking, and is a key player in regulation of the mucosal immune response. Zonulin is a marker of intestinal permeability. Its concentrations correlate significantly with total calorie-, protein-, carbohydrate intake. Zonulin plays a role in the pathogenesis of malabsorption in two forms of malnutrition (anorexia nervosa and celiac disease) and probably has an important role in the pathogenesis of wasting. During malnutrition, Zonulin levels may increase due to inflammation, infection, injury, or poor diet and enhance intestinal permeability which leads to paracellular passage of non-self-antigens (D) into the lamina propria (loose connective tissue in the mucosa), where these foreign antigens interact with the immune system. It has been shown that serum zonulin levels and other markers of barrier dysfunction is correlated with stunted growth in EED patients. (60)

In mice, it has been demonstrated that vitamin D deficiency (VDD), a common association in undernourished children, could lead to a significant upregulation in mRNA expression of the jejunum zonulin level with increase of serum level of zonulin. (60) Directly blocking zonulin by larazotide acetate (AT-1001), a first-in-class TJ regulator, is currently used in phase III clinical trials for celiac disease to improve intestinal barrier function. (61) The first randomized controlled phase IIb trial of an anti-inflammatory drug for environmental enteric dysfunction (EED) using aminosalicylate mesalazine proved its safety with modest reductions in several inflammatory markers compared to placebo. Before mesalazine treatment the IGF-1 concentration was negatively correlated with several inflammatory markers. With mesalazine treatment for 56 days, there was an increase in the IGF-1 level and decreases in the inflammatory marker concentrations that correlated with linear growth. (62)

3.3. Insulin and glucose homeostasis

Severe malnutrition predisposes children to develop either hypo- or hyperglycemia. During severe malnutrition, depleted glycogen stores and adipose tissue reserves have been correlated to hypoglycemia. In severely undernourished children, especially edematous forms hepatic glucose synthesis has been observed to be low. Hormonal changes with impaired insulin responses have been reported in severely undernourished children. Insulin responses to oral glucose and to a meal are strongly impaired in both kwashiorkor and marasmus, with low glucose clearance. Low insulin state decreases tissue and fat anabolism and permits the catabolic activity of other hormones to break down glycogen and fat to assure energy supply for vital organs and prevent hypoglycemia. Plasma insulin increases significantly during catch-up growth with nutritional rehabilitation. (63- 66) In addition, severe malnutrition, especially the edematous form, is associated with an impaired glucose absorption and decreased glucose absorption correlates with oxidative stress (infection and inflammation) in these children. (67,68)

In a meta-analysis review of 16 studies, Ledger et al reported that the prevalence of hypoglycemia in severely undernourished infants across studies based on of proportions was 9%. Meta-analysis results showed that hypoglycemia was associated with a higher chance of mortality during hospitalization in children with SAM (dys-adaptation). According to the GRADE evaluation, the certainty of the evidence for the prevalence of hypoglycemia was low and for hyperglycemia was very low. (68)

Recent findings have shown an important role for insulin in shaping the immune response during an infection. This includes the ability of insulin to modulate immune cell differentiation and polarization as well as the modulation of effector functions such as biocidal ROS production. On the other hand, both through a direct and indirect effects inflammatory mediators can control serum insulin levels. Therefore, low insulin status in severely undernourished children may adversely affect their immune response during an infection. (69)

3.4. Hypothalamic Pituitary adrenal (HPA) and GH (HPG) axes:

HPA Axis Cortisol increases markedly in children with malnutrition, especially those with infection. Hypercortisolemia causes muscle wasting both in vivo and in vitro by increasing protein breakdown and reducing protein synthesis. Metabolically, raised cortisol levels increase the availability of all fuel substrates by mobilization of glucose, free fatty acids, and amino acids from endogenous stores which reduces muscle mass and may enhance energy expenditure. The release of amino acids from muscle provides good substrate for gluconeogenesis during malnutrition which guards against hypoglycemia. (70) On the other hand, cortisol excess can adversely arrest bone growth in these children. (this spares energy for the major metabolic processes during malnutrition) (71,72)

In addition, glucocorticoids inhibit many of the initial events in inflammatory and immune responses. High cortisol inhibits the vasodilation and vascular permeability that occurs following inflammatory insult and decreases leukocyte emigration into inflamed sites, effects that require new protein synthesis. High cortisol represses transcription of many genes encoding pro-inflammatory cytokines and chemokines, cell adhesion molecules and key enzymes involved in the initiation and/or maintenance of the host inflammatory response. High cortisol can alter leukocyte distribution/trafficking, death/survival and, significantly, alter cellular differentiation programs, thus shaping the subsequent response. These different effects can be shared in the compromised immune and inflammation responses in undernourished infants. In fact, serum cortisol concentration was shown to be associated with severity and mortality in patients with pneumonia. (72-74)

3.5. Hypothalamic pituitary- growth hormone -IGF1 axis

An appropriate balance between the immune and GH-IGF systems is necessary for normal growth and immunity. In humans, GH is a critical factor to increase the rate of lipolysis during fasting. Growth hormone levels are high in children with malnutrition and represent an important means of mobilizing fat stores and maintaining euglycemia in states of under-nutrition. Yet, if the GH-IGF-I axis remained intact during states of nutritional deprivation, the elevated levels of GH would result in elevated IGF-I levels, leading to increased energy expenditure which is a clear disadvantage that compromises survival during malnutrition. Therefore, GH resistance, with an inability of GH to appropriately generate IGF-I production, is likely an adaptive mechanism to preserve calories during periods of under-nutrition. Protein deficiency not only results in GH resistance but also likely results in a state of end-organ resistance to IGF-I, with the result inhibiting growth. There appears to be a good adaptive mechanism where growth arrest spares energy necessary for survival. (72, 76) Zinc, magnesium, and vitamin B6 deficiencies that occur in many children with malnutrition have been associated with GH resistance and reduced IGF-I levels, although the mechanisms of each are unknown. (77)

Growth hormone and IGF-1 have important immunoregulatory effects. In a large cohort of undernourished children, infection and inflammation were associated with GH resistance, a low level of IGF-1, as well as with growth impairment. (78) In addition, GH and IGF-1 may act to protect the host from lethal bacterial infection by promoting the maturation of myeloid cells, stimulating phagocyte migration, priming them to produce superoxide anions and cytokines, and enhancing opsonic activity. Therefore, GH resistance and low IGF1 may compromise these immunoregulatory functions during malnutrition and infection. (72,78,79) Moreover, IGF-I plays an essential role in the growth, stimulation, proliferation, and function of T cells. IGF-I regulates various aspects of T-cell, B-cell, and monocyte function through its interactions with IGF-IR. IGF-I can prolong lymphocyte survival through activation of T cell Akt. (80)

IGFs depress proinflammatory cytokine signaling by increasing IL-10 secretion and via JNK and NF- κ B pathways. Therefore, in malnutrition IGF1 deficiency may negatively affect the functions of lymphocytes and predispose to infection as well as inflammation. Patients with inflammatory bowel disease (IBD) with low IGF1 have associated high inflammatory activity (ESR, CRP, and IL6). Suppression of inflammation by prednisolone or infliximab increased the IGF-1 levels and suppressed the inflammatory process. (80,81,82)

On the other hand, pro-inflammatory cytokines, which increase in severe forms of malnutrition especially those with infection, induce a dysregulation in GH-IGF axis and IGF system, both at central and peripheral levels. In the brain, inflammation/infection determines a dysregulation of GH secretion. (83)

In the liver, TNF- α , a pro-inflammatory cytokine can cause GH resistance mainly through downregulation of liver GH receptor expression. Additionally, the predominance of proinflammatory cytokines decreases IGF sensitivity by enhancing IGFBP production and by decreasing signaling through the (insulin receptor signaling) IRS/ Akt pathway. (83,84) During malnutrition, GH and IGF-1 resistance are also present in the growth plate. In these children and infants' abnormalities in IGF binding proteins (IGFBPs), with low IGFBP 3 levels, lead to a decline in IGF bioavailability. MicroRNAs (miRNAs) genes are known to regulate GH, IGF1, IGF2, and IGF1R in the context of body growth. During malnutrition, miRNAs targeting genes within the GH-IGF axis and IGF system are dysregulated. (83) The increase in local muscle cytokines produced during inflammation makes the muscle GH-resistant and reduces its own IGF-I production. Both decreased IGF-I production by muscle, and lower muscle sensitivity to the anabolic effects of IGF-I, may contribute to muscle wasting observed in response to severe malnutrition. (83,84) Myokines secreted by the skeletal muscle itself in response to inflammation have been implicated as autocrine and paracrine mediators of cachexia, as well as potential modulators of this debilitating condition. (85)

The first randomized controlled in the initial management of severely acutely undernourished children with environmental enteric dysfunction (EED) using aminosalicylate mesalazine proved its safety with modest reductions in several inflammatory markers compared to placebo. Before mesalazine treatment, the IGF-1 concentration was

negatively correlated with several inflammatory markers. With mesalazine treatment for 56 days, there was an increase in the IGF-1 level and decreases in the inflammatory marker concentrations that correlated with linear growth. (86)

3.6. Effect of Nutrition rehabilitation on hormonal changes and inflammatory markers

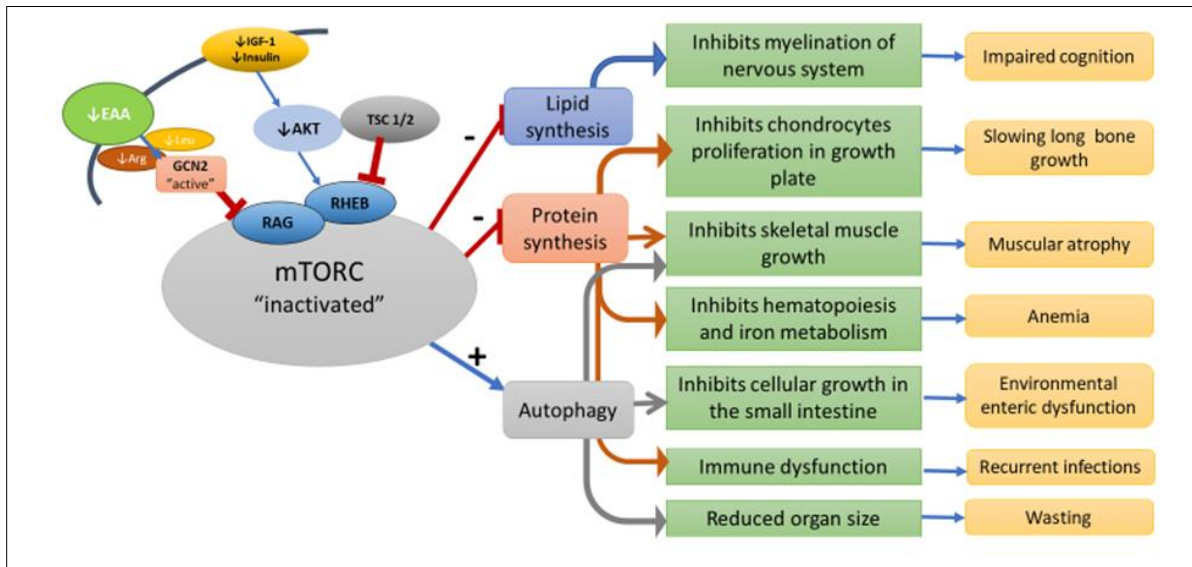


Figure 1 mTOR complex role in the pathogenesis of nutritional stunting and wasting in children.

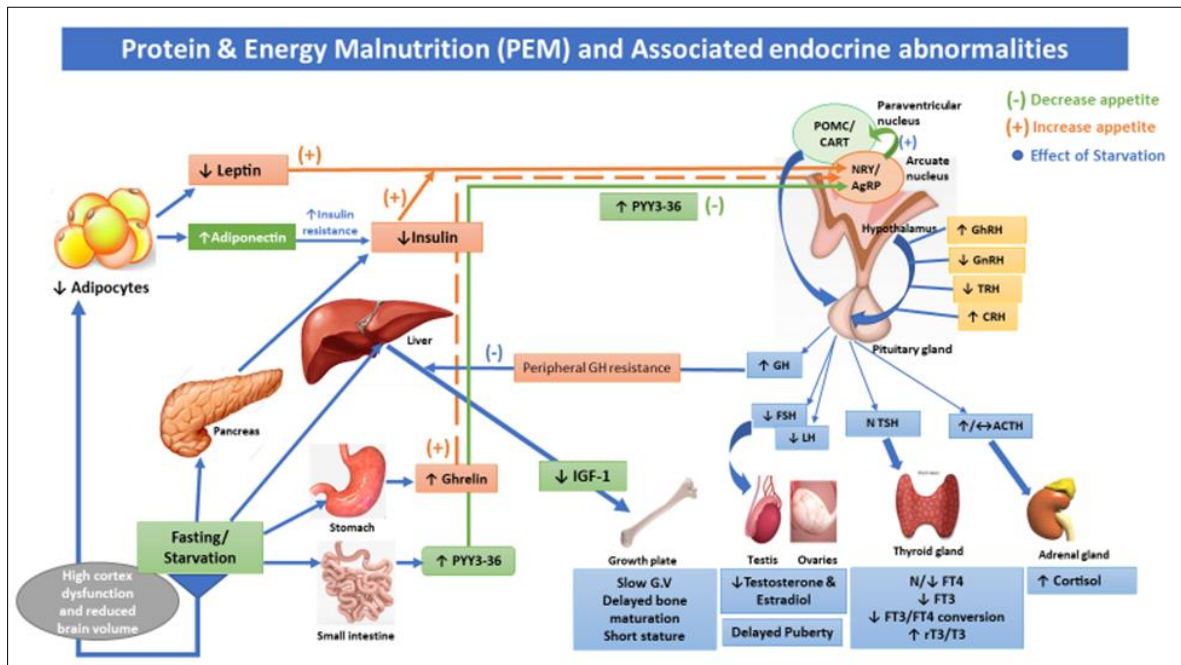


Figure 2 The interaction between malnutrition, central (hypothalamic-pituitary) and peripheral (IGF1, FT4, insulin) endocrine organs

Nutritional replacement in children with severe malnutrition can normalize the GH/IGF-I axis. Within two weeks of refeeding, IGF-I concentrations can double or triple and after 50 days of intensive inpatient nutritional therapy, basal GH and IGF-I levels are indistinguishable from controls. (57, 72, 87) Nutritional interventions are associated with early increases in IGF-I levels, even before changes in anthropometric measures are observed. In addition, the blunted increase in GH concentrations following arginine stimulation testing also resolves with treatment. In one study on undernourished infants and toddlers, insulin levels rose 50% during formula feeding, whereas leptin and IGF-I levels

increased nearly 3-fold. GH fell by 21% and cortisol levels also declined. In addition, after 2 weeks of formula feeding, leptin, and insulin levels increase significantly with refeeding while ghrelin, cortisol, and PYY decrease significantly. There were downward trends in almost all inflammatory cytokines. (57, 72, 87, 88) Therefore, it is understandable that all the consequences of low IGF1 (Metabolic – immune and growth) are correctable when proper nutrition restores the normal IGF1 level necessary for catch-up growth during rehabilitation.

4. Conclusion

In summary, it appears that in infants and children, the endocrine changes in response to malnutrition mediate an adaptation process that secures energy substrates to vital organs and prevents hypoglycemia through a catabolic process (high GH, high cortisol, and low insulin, IGF1, and leptin) involving lipolysis, gluconeogenesis, and muscle wasting. Meanwhile, this process impairs/stops linear growth (low IGF1) to spare energy. These endocrine changes however may have a negative effect on systemic immune and inflammatory responses to infections (high cortisol, low IGF1, and low insulin) which are commonly associated with malnutrition. However, on nutritional rehabilitation, all these endocrine changes are easily reverted to normal even before any change occurs in the anthropometric data. (Weight gain).

Compliance with ethical standards

Acknowledgments

We are grateful to our dietitians with whom we have had the pleasure to work to achieve nutritional rehabilitation of our malnourished and failure to thrive children (Maya Itani, Mona Shaat, and Doaa Yassin).

Disclosure of conflict of interest

There is no conflict of interest among authors.

Funding

This research did not receive any specific grant from funding agencies in the public, commercial, or not-for-profit sectors.

Author's Contribution

- Ashraf Tawfik Soliman, MD, Ph.D. FRCP: design of the work: Data collection, analysis and interpretation and drafting the article.
- Nada Mwafak Alaaraj, MD: Data collection, analysis and drawing the figures.
- Doaa Khater MD: shared in the drafting and revision of the article.
- Alan D. Rogol, MD Ph. D.: Data interpretation and critical revision from the endocrine point of view

References

- [1] Black RE, Cousens S, Johnson HL, et al. Global, regional, and national causes of child mortality in 2008: a systematic analysis. *Lancet*. 2010, 375(9730):1969–1987 [PubMed] [Google Scholar]
- [2] Black RE, Allen LH, Bhutta ZA, et al. Maternal and child undernutrition: global and regional exposures and health consequences. *Lancet*. 2008, 371(9608):243–260 [PubMed] [Google Scholar]
- [3] Lozano R, Mohsen N, Foreman K, et al. Global and regional mortality from 235 causes of death for 20 age groups in 1990 and 2010: a systematic analysis for the Global Burden of Disease Study 2010. *Lancet*. 2012, 380(9859):2095–2128
- [4] Dewey KG, Begum K. Long-term consequences of stunting in early life. *Matern Child Nutr*. 2011, 7:5.
- [5] Black RE, Allen LH, Bhutta ZA, et al. Maternal and child undernutrition 1- Maternal and child undernutrition: global and regional exposures and health consequences. *Lancet*. 2008, 371:243- 260.
- [6] Perkins JM, Subramanian SV, Davey Smith G, Özaltın E. Adult height, nutrition, and population health. *Nutr Rev*. 2016, 74(3):149-165. doi:10.1093/nutrit/nuv105

- [7] Soliman A, De Sanctis V, Alaaraj N, Ahmed S, Alyafei F, et al. Early and Long-term Consequences of Nutritional Stunting: From Childhood to Adulthood. *Acta Biomed.* 2021 Feb 16, 92(1):e2021168. doi: 10.23750/abm.v92i1.11346. PMID: 33682846, PMCID: PMC7975963.
- [8] Frohman LA, Neuroendocrinology, Editor(s): Larry R. Squire, *Encyclopedia of Neuroscience*, Academic Press, 2009, Pages 351-354, ISBN 9780080450469, <https://doi.org/10.1016/B978-008045046-9.01184-0>.
- [9] Procaccini C, Pucino V, De Rosa V, Marone G, Matarese G. Neuro-endocrine networks controlling immune system in health and disease. *Front Immunol.* 2014, 5:143. Published 2014 Apr 7. doi:10.3389/fimmu.2014.00143
- [10] Bou Nemer L, Shi H, Carr BR, Word RA, Bukulmez O. Effect of Body Weight on Metabolic Hormones and Fatty Acid Metabolism in Follicular Fluid of Women Undergoing In Vitro Fertilization: A Pilot Study. *Reprod Sci.* 2019 Mar, 26(3):404-411. doi: 10.1177/1933719118776787. Epub 2018 May 20. PMID: 29779472.
- [11] Zuure WA, Roberts AL, Quennell JH, Anderson GM. Leptin signaling in GABA neurons, but not glutamate neurons, is required for reproductive function. *J Neurosci.* 2013, 33(45):17874-17883. doi:10.1523/JNEUROSCI.2278-13.2013.
- [12] Ibrahim Abdalla MM. Ghrelin - Physiological Functions and Regulation. *Eur Endocrinol.* 2015, 11(2):90-95. doi:10.17925/EE.2015.11.02.90
- [13] Yoo ES, Yu J, Sohn JW. Neuroendocrine control of appetite and metabolism. *Exp Mol Med.* 2021 Apr, 53(4):505-516. doi: 10.1038/s12276-021-00597-9. Epub 2021 Apr 9. PMID: 33837263, PMCID: PMC8102538.
- [14] Caulfield LE, de Onis M, Blossner M, Black RE. Undernutrition as an underlying cause of child deaths associated with diarrhea, pneumonia, malaria, and measles. *Am J Clin Nutr.* 2004, 80(1):193-198
- [15] Ibrahim MK, Zambruni M, Melby CL, Melby PC. Impact of Childhood Malnutrition on Host Defense and Infection. *Clin Microbiol Rev.* 2017, 30(4):919-971.
- [16] Peter Katona, Judit Katona-Apte. The Interaction between Nutrition and Infection, *Clinical Infectious Diseases*, Volume 46, Issue 10, 15 May 2008, 1582-1588.
- [17] Taylor HL, Keys A. Adaptation to caloric restriction. *Science.* 1950, 112(2904):215-19.
- [18] Cahill GF., Jr Starvation in man. *N Engl J Med.* 1970, 282(12):668-675.
- [19] Lunn PG, Whitehead RG, Coward WA. Two pathways to kwashiorkor? *Trans R Soc Trop Med Hyg.* 1979, 73(4):438-444
- [20] Whitehead RG, Lunn PG. Endocrines in protein-energy malnutrition. *Proc Nutr Soc.* 1979, 38(1):69-76.
- [21] Jahoor F, Badaloo A, Reid M, Forrester T. Protein metabolism in severe childhood malnutrition. *Ann Trop Paediatr.* 2008, 28(2):87-101
- [22] Ulrich-Lai YM, Ryan KK. Neuroendocrine circuits governing energy balance and stress regulation: functional overlap and therapeutic implications. *Cell Metab.* 2014, 19(6):910-925. doi:10.1016/j.cmet.2014.01.020
- [23] Ulrich-Lai YM, Herman JP. Neural regulation of endocrine and autonomic stress responses. *Nat Rev Neurosci.* 2009, 10(6):397-409. doi:10.1038/nrn2647
- [24] Rønnestad I, Gomes AS, Murashita K, Angotzi R, Jönsson E, Volkoff H. Appetite-Controlling Endocrine Systems in Teleosts. *Front Endocrinol (Lausanne).* 2017 Apr 18, 8:73. doi: 10.3389/fendo.2017.00073. PMID: 28458653, PMCID: PMC5394176.
- [25] Delgado TC. Glutamate and GABA in Appetite Regulation. *Front Endocrinol (Lausanne).* 2013, 4:103.
- [26] Quarta C, Claret M, Zeltser LM, Williams KW, Yeo GSH, et al POMC neuronal heterogeneity in energy balance and beyond: an integrated view. *Nat Metab.* 2021 Mar, 3(3):299-308.
- [27] Soliman GA. The role of mechanistic target of rapamycin (mTOR) complexes signaling in the immune responses. *Nutrients.* 2013 Jun 19, 5(6):2231-57.
- [28] Sengupta S, Peterson TR, Sabatini DM. Regulation of the mTOR complex 1 pathway by nutrients, growth factors, and stress. *Mol Cell.* 2010, 40(2):310-322. doi: 10.1016/j.molcel.2010.09.026
- [29] Phornphutkul C, Wu KY, Auyeung V, Chen Q, Gruppuso PA. mTOR signaling contributes to chondrocyte differentiation. *Dev Dyn.* 2008, 237:702-712.

- [30] Yoon MS. mTOR as a Key Regulator in Maintaining Skeletal Muscle Mass. *Front Physiol.* 2017, 8:788. Published 2017 Oct 17. doi:10.3389/fphys.2017.00788
- [31] Blouet C., Ono H., Schwartz G.J. Mediobasal Hypothalamic p70 S6 Kinase 1 Modulates the Control of Energy Homeostasis. *Cell Metab.* 2008, 8:459–467. doi: 10.1016/j.cmet.2008.10.004.
- [32] Mao Z, Zhang W. Role of mTOR in Glucose and Lipid Metabolism. *Int J Mol Sci.* 2018, 19(7):2043. Published 2018 Jul 13. doi:10.3390/ijms19072043
- [33] Hu F., Xu Y., Liu F. Hypothalamic roles of mTOR complex I: Integration of nutrient and hormone signals to regulate energy homeostasis. *Am. J. Physiol.-Endocrinol. Metab.* 2016, 310:E994–E1002
- [34] Yu JH, Kim MS. Molecular mechanisms of appetite regulation. *Diabetes Metab J.* 2012, 36:391–398.
- [35] Muta K, Morgan DA, Rahmouni K. The role of hypothalamic mTORC1 signaling in insulin regulation of food intake, body weight, and sympathetic nerve activity in male mice. *Endocrinology.* 2015, 156:1398–1407.
- [36] Geissler EK. The influence of mTOR inhibitors on immunity and the relationship to post-transplant malignancy. *Transplant Res.* 2013, 2(Suppl 1):S2. doi:10.1186/2047-1440-2-S1-S2
- [37] Akib RD, Aminuddin A, Hamid F, Prihantono P, Bahar B, et al. Leptin levels in children with malnutrition. *Gac Sanit.* 2021, 35 Suppl 2:S278-S280.
- [38] Soliman AT, ElZalabany MM, Salama M, Ansari BM. Serum leptin concentrations during severe protein-energy malnutrition: correlation with growth parameters and endocrine function. *Metabolism.* 2000 , 49(7):819-25.
- [39] Büyükgebiz B, Oztürk Y, Yilmaz S, Arslan N. Serum leptin concentrations in children with mild-to-moderate protein-energy malnutrition. *Pediatr Int.* 2003 , 45(5):550-4.
- [40] Haspolat K, Ece A, Gürkan F, Atamer Y, Tutanç M, Yolbaş I. Relationships between leptin, insulin, IGF-1 and IGFBP-3 in children with energy malnutrition. *Clin Biochem.* 2007,40(3-4):201-5.
- [41] Perry RJ, Shulman GI. The Role of Leptin in Maintaining Plasma Glucose During Starvation. *Postdoc J.* 2018, 6(3):3-19. doi: 10.14304/surya.jpr.v6n3.2
- [42] Farooqi IS, Wangensteen T, Collins S, Kimber W, Matarese G, et al. Clinical and molecular genetic spectrum of congenital deficiency of the leptin receptor. *N Engl J Med.* 2007, 18, 356(3):237-47.
- [43] Fernández-Riejos P, Najib S, Santos-Alvarez J, et al. Role of leptin in the activation of immune cells. *Mediators Inflamm.* 2010, 2010:568343. doi:10.1155/2010/568343
- [44] Maurya R, Bhattacharya P, Dey R, Nakhasi HL. Leptin Functions in Infectious Diseases. *Front Immunol.* 2018, 9:2741. Published 2018 Nov 26. doi:10.3389/fimmu.2018.02741
- [45] Osório, J. Low leptin levels linked with mortality in severely malnourished children. *Nat Rev Endocrinol* 10, 252 (2014). <https://doi.org/10.1038/nrendo.2014.30>
- [46] Bartz S, Mody A, Hornik C, et al. Severe acute malnutrition in childhood: hormonal and metabolic status at presentation, response to treatment, and predictors of mortality. *J Clin Endocrinol Metab.* 2014, 99(6):2128-2137. doi:10.1210/jc.2013-4018
- [47] Khoramipour K, Chamari K, Hekmatikar AA, Ziyaiyan A, Taherkhani S, et al. Adiponectin: Structure, Physiological Functions, Role in Diseases, and Effects of Nutrition. *Nutrients.* 2021 Apr 2, 13(4):1180. doi: 10.3390/nu13041180. PMID: 33918360, PMCID: PMC8066826
- [48] Nassar MF, Badawy NB, Ezz El-Arab S, El-Batrawy SR, Kamal M.A. Adiponectin level in Protein Energy Malnutrition and its Role in Predicting the Disease Severity. *Journal of American Science*, 2010, 6(12)
- [49] Pannacciulli N, Vettor R, Milan G, Granzotto, M, Catucci A, Federspil G, et al. Anorexia nervosa is characterized by increased adiponectin plasma levels and reduced non-oxidative glucose metabolism. *J Clin Endocrinol Metab.* 2003, 88: 1748-1752.
- [50] Liu Y, Sweeney G. Adiponectin action in skeletal muscle. *Best Pract Res Clin Endocrinol Metab.* 2014 Jan, 28(1):33-41.
- [51] Abou-Samra M, Selvais CM, Dubuisson N, Brichard SM. Adiponectin and Its Mimics on Skeletal Muscle: Insulin Sensitizers, Fat Burners, Exercise Mimickers, Muscling Pills ... or Everything Together? *Int J Mol Sci.* 2020, 9, 21(7):2620.

- [52] Luo Y, Liu M. Adiponectin: a versatile player of innate immunity. *J Mol Cell Biol.* 2016, 8(2):120-128. doi:10.1093/jmcb/mjw012
- [53] De Silva A, Bloom SR. Gut hormones and appetite control: a focus on PYY and GLP-1 as therapeutic targets in obesity. *Gut Liver.* 2012, 6(1):10–20
- [54] Vijayakumar A, Novosyadlyy R, Wu Y, Yakar S, LeRoith D. Biological effects of growth hormone on carbohydrate and lipid metabolism. *Growth Horm IGF Res.* 2010, 20(1):1–7
- [55] Yi CX, Heppner KM, Kirchner H, et al. The GOAT-ghrelin system is not essential for hypoglycemia prevention during prolonged calorie restriction. *PLoS One.* 2012, 7:e32100.
- [56] Pradhan G, Samson SL, Sun Y. Ghrelin: much more than a hunger hormone. *Curr Opin Clin Nutr Metab Care.* 2013, 16(6):619-624. doi:10.1097/MCO.0b013e328365b9be
- [57] Welsh FK, Farmery SM, MacLennan K, Sheridan MB, Barclay GR, et al. Gut barrier function in malnourished patients. *Gut.* 1998 Mar, 42(3):396-401. doi: 10.1136/gut.42.3.396.
- [58] Guerrant RL, Leite AM, Pinkerton R, Medeiros PH, Cavalcante PA, et al. Biomarkers of Environmental Enteropathy, Inflammation, Stunting, and Impaired Growth in Children in Northeast Brazil. *PLoS One* 2016, 11: e0158772, PMID:27690129.
- [59] Yeung CY, Chiang Chiau JS, Cheng ML, Chan WT, Jiang CB, et al. Effects of Vitamin D-Deficient Diet on Intestinal Epithelial Integrity and Zonulin Expression in a C57BL/6 Mouse Model. *Front Med (Lausanne).* 2021 Aug 3, 8:649818.
- [60] Sturgeon, C. & Fasano, A. Zonulin, a regulator of epithelial and endothelial barrier functions, and its involvement in chronic inflammatory diseases. *Tissue Barriers* 2016, 4, e1251384.
- [61] Bartels RH, Chimwezi E, Watson V, Pei L, Potani I, et al. Hypoallergenic and anti-inflammatory feeds in children with complicated severe acute malnutrition: an open randomised controlled 3-arm intervention trial in Malawi. *Sci Rep.* 2019, 19, 9(1):2304.
- [62] Bandsma RH, Mendel M, Spoelstra MN, Reijngoud DJ, Boer T, et al. Mechanisms behind decreased endogenous glucose production in malnourished children. *Pediatr Res.* 2010, 68(5):423–8.
- [63] Spoelstra MN, Mari A, Mendel M, Senga E, van Rheenen P, et al. Kwashiorkor and marasmus are both associated with impaired glucose clearance related to pancreatic β -cell dysfunction. *Metabolism.* 2012 Sep, 61(9):1224-30. doi: 10.1016/j.metabol.2012.01.019. Epub 2012 Mar 3. PMID: 22386944.
- [64] Robinson HM, Seakins A. Fasting pancreatic glucagon in Jamaican children during malnutrition and subsequent recovery. *Pediatr Res.* 1982 Dec, 16(12):1011-5. doi: 10.1203/00006450-198212000-00008. PMID: 6818514.
- [65] Soliman AT, Aref MK, Hassan AE. Hormonal changes in protein energy malnutrition. *Indian J Pediatr.* 1988 Jul-Aug, 55(4):465-9. doi: 10.1007/BF02868424. PMID: 3139559.
- [66] Bandsma RH, Spoelstra MN, Mari A, Mendel M, van Rheenen PF, Senga E, et al. Impaired glucose absorption in children with severe malnutrition. *J Pediatr.* 2011 Feb, 158(2):282-7.e1. doi: 10.1016/j.jpeds.2010.07.048. Epub 2010 Sep 16. PMID: 20843523.
- [67] Ivanov AV, Bartosch B, Isaguliantz MG. Oxidative Stress in Infection and Consequent Disease. *Oxid Med Cell Longev.* 2017, 2017:3496043. doi:10.1155/2017/3496043
- [68] Elizabeth Ledger, Philliness Prisca Harawa, Allison I Daniel, Toby Candler, et al. Dysglycemia in Children with Severe Acute Malnutrition: A Systematic Review and Meta-Analysis, *Advances in Nutrition*, Volume 12, Issue 3, May 2021, Pages 959–968,
- [69] van Niekerk G, Christowitz C, Conradie D, Engelbrecht AM. Insulin as an immunomodulatory hormone. *Cytokine Growth Factor Rev.* 2020 Apr, 52:34-44. doi: 10.1016/j.cytogfr.2019.11.006. Epub 2019 Dec 3. PMID: 31831339.
- [70] Menconi M, Fareed M, O'Neal P, Poylin V, Wei W, et al. Role of glucocorticoids in the molecular regulation of muscle wasting. *Crit Care Med.* 2007, 35(9 Suppl):S602-8.
- [71] Soliman AT, Hassan AE, Aref MK, Hintz RL, Rosenfeld RG, et al. Serum insulin-like growth factors I and II concentrations and growth hormone and insulin responses to arginine infusion in children with protein-energy malnutrition before and after nutritional rehabilitation. *Pediatr Res.* 1986, 20(11):1122–1130.
- [72] Perretti M, Ahluwalia A. The microcirculation and inflammation: site of action for glucocorticoids. *Microcirculation.* 2000 Jun, 7(3):147-61.

- [73] Coutinho AE, Chapman KE. The anti-inflammatory and immunosuppressive effects of glucocorticoids, recent developments and mechanistic insights. *Mol Cell Endocrinol.* 2011, 335(1):2-13. doi: 10.1016/j.mce.2010.04.005
- [74] Kolditz, M., Höffken, G., Martus, P. et al. Serum cortisol predicts death and critical disease independently of CRB-65 score in community-acquired pneumonia: a prospective observational cohort study. *BMC Infect Dis* 12, 90 (2012). <https://doi.org/10.1186/1471-2334-12-90>
- [75] Fazeli PK, Klibanski A. Determinants of GH resistance in malnutrition. *J Endocrinol.* 2014, 220(3): R57-R65. Published 2014 Jan 27. doi:10.1530/JOE-13-0477
- [76] Caputo M, Pigni S, Agosti E, et al. Regulation of GH and GH Signaling by Nutrients. *Cells.* 2021, 10(6):1376. Published 2021 Jun 2. doi:10.3390/cells10061376
- [77] DeBoer MD, Scharf RJ, Leite AM, Ferrer A, Havt A, Pinkerton R, et al. Systemic inflammation, growth factors, and linear growth in the setting of infection and malnutrition. *Nutrition.* 2017, 33:248-253.
- [78] Saito H, Inoue T, Fukatsu K, Ming-Tsan L, Inaba T, et al. Growth hormone and the immune response to bacterial infection. *Horm Res.* 1996, 45(1-2):50-4.
- [79] Smith TJ. Insulin-like growth factor-I regulation of immune function: a potential therapeutic target in autoimmune diseases? *Pharmacol Rev.* 2010, 62(2):199-236. doi:10.1124/pr.109.002469
- [80] Basu S, Rajakaruna S, Menko AS. Insulin-like growth factor receptor-1 and nuclear factor κ B are crucial survival signals that regulate caspase-3-mediated lens epithelial cell differentiation initiation. *J Biol Chem.* 2012 Mar 9, 287(11):8384-97.
- [81] O'Connor JC, McCusker RH, Strle K, Johnson RW, Dantzer R, et al. Regulation of IGF-I function by proinflammatory cytokines: at the interface of immunology and endocrinology. *Cell Immunol.* 2008, 252(1-2):91-110.
- [82] Cirillo F, Lazzeroni P, Sartori C, Street ME. Inflammatory Diseases and Growth: Effects on the GH-IGF Axis and on Growth Plate. *Int J Mol Sci.* 2017, 18(9):1878.
- [83] Thissen JP: How Proinflammatory Cytokines May Impair Growth and Cause Muscle Wasting. *Horm Res* 2007, 67(suppl 1):64-70.
- [84] Webster JM, Kempen LJAP, Hardy RS, Langen RCJ. Inflammation and Skeletal Muscle Wasting During Cachexia. *Front Physiol.* 2020,19, 11:597675.
- [85] Jones, K.D., Hünten-Kirsch, B., Laving, A.M. et al. Mesalazine in the initial management of severely acutely malnourished children with environmental enteric dysfunction: a pilot randomized controlled trial. *BMC Med* 12, 133 (2014).
- [86] Kouanda S, Doulogou B, De Coninck V, Habimana L, Sondo B, Tonglet R, Ketelslegers J, Robert A. Insulin Growth Factor-I in Protein-Energy Malnutrition during Rehabilitation in Two Nutritional Rehabilitation Centres in Burkina Faso. *J Trop Med.* 2009, 2009:832589.
- [87] Caregato L, Favaro A, Santonastaso P, Alberino F, Di Pascoli L, et al A. Insulin-like growth factor 1 (IGF-1), a nutritional marker in patients with eating disorders. *Clin Nutr.* 2001, 20(3):251–257.