

eISSN: 2581-9615 CODEN (USA): WJARAI Cross Ref DOI: 10.30574/wjarr Journal homepage: https://wjarr.com/

WJARR	HISSN:3581-8615 CODEN (UBA): HUARAI		
W	JARR		
World Journal of			
Advanced			
Research and			
Reviews			
	World Journal Series INDIA		
Check for undates			

(RESEARCH ARTICLE)

Evaluation of anthelmintic activity of *dryopteris filix*-mas using earthworm

Dharamveer Prajapati *, Arvind Singh Jadon and Kapil Purohit

Gurukul Institute of Pharmaceutical Science and Research, Gwalior, Madhya Pradesh, India-464001.

World Journal of Advanced Research and Reviews, 2022, 16(01), 886-893

Publication history: Received on 19 September 2022; revised on 27 October 2022; accepted on 30 October 2022

Article DOI: https://doi.org/10.30574/wjarr.2022.16.1.1093

Abstract

Evaluation of anthelmintic activity of a leaves extracts of *Dryopteris filix*-mas against earthworm with Preliminary work as physical characteristics. Finally extraction of defatted *Dryopteris filix*-mas extract was done with hydro-alcoholic solvent and % yield was found to be 6.55% w/w and their characteristics are reported. Results obtained from qualitative chemical tests are tabulated. Total alkaloid content was calculated as atropine equivalent mg/100 mg using the equation based on the calibration curve: Y=0.007X+0.007, R2=0.999, where X is the Atropine equivalent (AE) and Y is the absorbance. Total phenol content was expressed as mg/100 mg of gallic acid equivalent of dry extract sample using the equation obtained from the calibration curve: Y = 0.017X+0.017, R2= 0.998, where X is the gallic acid equivalent (GAE) and Y is the absorbance. The present findings indicated that the usefulness of the hydroalcoholic extract of leaves of *Dryopteris filix*-mas in earthworm. This study suggested, hydroalcoholic extract of leaves of *Dryopteris filix*-mas dose-dependently produced anthelmintic activity.

Keywords: Anthelmintic; Dryopteris filix-mas; Earthworm; Hydroalcoholic

1. Introduction

Nature has provided a complete store-house of remedies to cure all ailments of mankind and its related diseases¹. Charaka made fifty groups of ten herbs each of them sufficient for an ordinary physician's need. Sushruta arranged 760 herbs in 7 distinct sets based on some of their common properties². Most diseases caused by helminths are chronic, debilitating in nature, they probably cause more morbidity, greater economic and social deprivation among humans and animals than any other parasites³. It is not only limited to tropical and subtropical countries but is also to endemic in many regions because of poor sanitation, poor family hygiene, malnutrition and crowded living condition⁴. Potent anthelminitics are available today, and treatment is frequently done by using different types of drugs⁵. However the high costs of modern anthelminitics have limited effective control of the parasites⁶. In some cases, wide spread use of low quality anthelminitics are used for the development of resistance and hence causes reduction in use of anthelminitics⁷. Recently the use of anthelminitics produces toxicity in human beings. Hence the development and discovery of new substances acting as anthelminitics are being derived through plants which are considered to be the best source of bioactive substances⁸. Various plants were used in veneral diseases, leucorrhoea, dysentery, dysuria and fever^{9,10}. Anthelminitics are those drugs that are used in expelling out the worms that are parasitic in nature by either stunning them or by killing them. They are also known as vermifuges or vermicides¹¹.

Evaluations of anthelmintic activity of *Dryopteris filix*-mas drug was carried out in artificial laboratory conditions by using the earthworms (*Lumbricus terrestris*). The earthworms are ecofriendly for decomposing organic materials, feeding upon undecayed leaves and other plant materials, more geophagous.

* Corresponding author: Dharamveer Prajapati

Gurukul Institute of Pharmaceutical Science and Research, Gwalior, Madhya Pradesh, India- 464001.

Copyright © 2022 Author(s) retain the copyright of this article. This article is published under the terms of the Creative Commons Attribution Liscense 4.0.

2. Material and methods

2.1. Preliminary Work

The plant has been selected on the basis on Availability of plant material and its wide geographical distribution globally also economic plant. Leaves of *Dryopteris filix*-mas were collected from local areas and authenticated. Fresh leaves drying were carried out in sun but under the shade. They were pulverized to make coarse powder. The coarse powder of fruit was passed through sieve No. 18 to maintain uniformity. Dried leaves were preserved in plastic bags and closed tightly and powdered as per the requirements.

2.2. Method of Extraction

Following procedure was adopted for the preparation of extracts from the shade dried and powdered herbs:

2.2.1. Defatting of plant material

Leaves of *Dryopteris filix*-mas were shade dried at room temperature. 70.50 gram of dried leaves was coarsely powdered and subjected to extraction with petroleum ether by maceration. The extraction was continued till the defatting of the material had taken place.

2.2.2. Extraction by maceration process

Defatted leaves of *Dryopteris filix*-mas were extracted with hydroalcoholic solvent (Ethanol: Water:75:25) using maceration process (48 hrs). The extract was evaporated above their boiling points. Finally, the percentage yields were calculated of the dried extracts.

2.2.3. Determination of percentage yield

The percentage yields of each extract were calculated by using following formula:

Percentage Yield =
$$\frac{\text{Weight of Extract}}{\text{Weight of powdered drug}} X 100$$

2.3. Qualitative Evaluation

Phytochemical tests were done as per the standard methods described for detection of alkaloids, carbohydrates, Saponins, phenols, flavonoids, proteins and diterpenes.

2.3.1. Quantitative studies of bioactive constituents

Estimation of total alkaloids content¹²

The plant extracts (1 mg) was dissolved in methanol, added 1ml of 2 N HCl and filtered. This solution was transferred to a separating funnel, 5 ml of bromocresol green solution and 5 ml of phosphate buffer were added. The mixture was shaken with 1, 2, 3 and 4 ml chloroform by vigorous shaking and collected in a 10-ml volumetric flask and diluted to the volume with chloroform. A set of reference standard solutions of atropine (40, 60, 80, 100 and 120 μ g/ml) were prepared in the same manner as described earlier. The absorbance for test and standard solutions were determined against the reagent blank at 470 nm with an UV/Visible spectrophotometer. The total alkaloid content was expressed as mg of AE/100 mg of extract.

Estimation of total phenol content13

The total phenol content of the extract was determined by the modifiedfolin-ciocalteu method. 10 mg Gallic acid was dissolved in 10 ml methanol, various aliquots of 10- 50μ g/ml was prepared in methanol. 10 mg of dried extract was dissolved in 10 ml methanoland filter. Two ml (1 mg/ml) of this extract was for the estimation of phenol. 2 ml of extract and each standard was mixed with 1 ml of Folin-Ciocalteu reagent (previously diluted with distilled water 1:10 v/v) and 1 ml (7.5g/l) of sodium carbonate. The mixture was vortexes for 15s and allowed to stand for 10min for color development. The absorbance was measured at 765 nm using a spectrophotometer.

2.4. In-vitro Evaluation of Anthelmintic Activity with Earthworm

2.4.1. Preparation of Drug Solutions

The standard drug albendazole was received from Lee Pharma, Hyderabad and test samples of hydroalcoholic extract of *Dryopteris filix*-mas prepared in collage Laboratory. Albendazole and test drugs were prepared as 12.5 mg/ml, 25 mg/ml, 50 mg/ml, 100 mg/ml, and 200 mg/ml concentrations using water and ethanol as solvents, respectively.

2.4.2. Collection of Earthworms

Earthworms were collected locally from compost plant situated in a nursery of Gwalior. The average size of worms was 5–8 cm. Earthworms were identified and authenticated. The anthelmintic activity was carried out as per the method described elsewhere. The assay was performed *in-vitro* using adult earthworms owing to their anatomical and physiological resemblance with the intestinal round worms, parasites of human beings for preliminary evaluation of anthelmintic activity. A concentration of standard drug (albendazole) and synthetic test sample extract was prepared as described earlier.

2.4.3. Evaluation of Anthelmintic Activity Using Earthworms

The anthelmintic activity was performed according to the method of Ghosh¹⁴ et al. Earthworms, each of average length of 6 cm, were placed in Petri dishes containing 2 ml of various drug concentrations, 12.5 mg/ml, 25 mg/ml, 50 mg/ml, 100 mg/ml, and 200 mg/ml of solutions. Albendazole solution was used as reference standard drug and distilled water as control. The worms were observed for the motility after incubating at 37°C. This was done after pouring the Petri dishes content in the wash basin and allowing the worms to move freely. By tapping the end of each worm with the index finger and applying a bit of pressure, the worms that were alive showed motility and those dead were nonmotile. The motile worms were returned to the respective Petri dishes containing drug solutions and the incubation process was carried out again. In the control, the worms were viable for at least twelve days, which is similar to the findings. The time taken for paralysis, motility activity of any sort and death time of worms were observed and recorded after ascertaining that the worms did not move neither when shaken vigorously nor when dipped in warm water (50 °C).

3. Results

3.1. Physical Characteristics of Extract

Finally extraction of defatted *Dryopteris filix*-mas extract was done with ethanol solvent and % yield was found to be 6.55% w/w and their characteristics are reported in table below

Table 1 Physical characteristics of extract

Extract	Consistency	Colour	Odour	% Yield
Hydroalcoholic	Solid	Dark Brown	Pungent	6.55

3.2. Qualitative Chemical Test

Results obtained from qualitative chemical tests are tabulated in table below

Table 2 Qualitative chemical tests of extract of Dryopteris filix-mas

S. No.	Bioactive constituents	Hydroalcoholic extract
1	Alkaloids	+ ve
2	Carbohydrates	+ ve
3	Glycosides	- ve
4	Saponins	- ve
5	Phenols	+ ve
6	Flavonoids	+ ve
7	Proteins	- ve
8	Diterpenes	- ve

⁽⁺ ve) - Present, (- ve) - Absent

3.3. Quantitative study

3.3.1. Estimation of total alkaloid content (TAC)

Total alkaloid content was calculated as atropine equivalent mg/100 mg using the equation based on the calibration curve: Y=0.007X+ 0.007, R²=0.999, where X is the Atropine equivalent (AE) and Y is the absorbance.



Figure 1 Graph of calibration curve of Atropine

3.3.2. Estimation of total phenol content (TPC)



Figure 2 Graph of Calibration curve of Gallic acid

Table 3 Estimation of total alkaloids and phenol content

Total alkaloids content	Total phenol content	
0.869 mg/100 mg	0.574 mg/100 mg	



Figure 3 Graph of total alkaloids and phenol content

Total phenol content was expressed as mg/100 mg of gallic acid equivalent of dry extract sample using the equation obtained from the calibration curve: Y = 0.017X+0.017, $R^2 = 0.998$, where X is the gallic acid equivalent (GAE) and Y is the absorbance.

3.4. In-vitro Evaluation of Anthelmintic Activity with Earthworm

Table 4 Anthelmintic activity of Albendazole (SD) and extract in earthworms

Concentration of drug sample (mg/ml)	Albandazole (standard) treated		Extract treated	
	Time of Paralysis (sec) Mean ± SEM	Time of Death (sec) Mean ± SEM	Time of Paralysis(sec) Mean ± SEM	Time of Death (sec) Mean ± SEM
12.5	989 ± 15	1304 ± 12	1154 ± 19	1473 ± 13
25	858 ± 23	1107 ± 17	964 ± 14	1246 ± 07
50	758 ± 12	1067 ± 21	823 ± 31	1143 ± 17
100	534 ± 26	783 ± 08	614 ± 10	924 ± 15
200	394 ± 17	635 ± 02	501 ±16	784 ± 35



Figure 4 Comparison of Paralysis Time



Figure 5 Comparison of Death Time



Figure 6 Extracts treated earthworms

4. Discussion

Evaluation of anthelmintic activity of a leaves extracts of *Dryopteris filix*-mas against earthworm with Preliminary work as physical characteristics. It was observed that leaves were Green, small ovate and smooth. The leaves *Dryopteris filix*-mas were dried under shade in laboratory. They were pulverized to make coarse powder. The coarse powder of fruit was passed through sieve No. 18 to maintain uniformity and stored in cool and dry place for further study. Powders of leaves *Dryopteris filix*-mas was subjected for extraction, hydroalcoholic solvent (Ethanol:Water:75:25) using maceration process (48 hrs), while petroleum ether was used for defatting of the waxy materials. The final yield with hydroalcoholic solvent was found 6.55%. Further obtained leaves extract was subjected for phytochemical analysis found that leaves extract was rich source of alkaloids, carbohydrate, phenol and flavonoids. Total alkaloid content was calculated 0.869 mg/100 mg as atropine equivalent and Total phenol content was 0.574 mg/100 mg expressed as gallic acid equivalent of dry extract sample.

In-vitro anthelmintic activity of leaves of *Dryopteris filix*-mas was performed by preparation of Albendazole and test drugs were prepared as 12.5 mg/ml, 25 mg/ml, 50 mg/ml, 100 mg/ml, and 200 mg/ml concentrations using water and ethanol as solvents, respectively. Earthworms were collected locally from compost plant situated in a nursery of Gwalior. The average size of worms was 5–8 cm. Earthworms, each of average length of 6 cm, were placed in Petri dishes containing 2 ml of various drug concentrations, 12.5 mg/ml, 25 mg/ml, 50 mg/ml, 100 mg/ml, and 200 mg/ml of solutions. Albendazole solution was used as reference standard drug and distilled water as control. The worms were observed for the motility after incubating at 37°C. This was done after pouring the Petri dishes contain in the wash basin and allowing the worms to move freely. By tapping the end of each worm with the index finger and applying a bit of pressure, the worms that were alive showed motility and those dead were nonmotile. The motile worms were returned to the respective Petri dishes containing drug solutions and the incubation process was carried out again. In the control, the worms were viable for at least twelve days, which is similar to the findings. The time taken for paralysis, motility activity of any sort and death time of worms were observed and record after ascertaining that the worms did not move neither when shaken vigorously nor when dipped in warm water (50°C).

In concentration of 12.5 mg/ml worms were paralyzed in 1154sec while dead in 1473sec. In concentration of 25 mg/ml worms were paralyzed in 964 sec while dead in 1246sec. In concentration of 50 mg/ml worms were paralyzed in 823sec while dead in 1143sec. In concentration of 100 mg/ml worms were paralyzed in 614sec while dead in 924sec. In concentration of 200 mg/ml worms were paralyzed in 501sec while dead in 784sec. Results were evaluated and

concluded that the potency of extract was increased with increasing of dose or concentration of extract. Therefore, it can be concluded that worms can be used successfully for the anthelmintic activity study as it is easy, prominent, an adaptable to laboratory conditions, and reproducible method in all aspects such as equal age, size and weight of the worms.

This experiment was to provide natural environment to the worms and was used for evaluating the effect different doses of the drugs on the viability of the preparasitic stages of the helminthics. In conclusion the adaptable factors known to us have influence on the anthelmintic activity of the drugs. Moreover, there was a need for an alternative method apart from the conventional method to justify anthelmintic studies in laboratory investigations. Further studies have to follow up the improved new methodology in evaluating the anthelmintic activity for any drug with potential role in worm infections.

5. Conclusion

In the last few decades eco-friendly, bio-friendly, cost effective and relatively safe, plant-based medicines have moved from the fringe to the main stream with the increased research in the field of traditional medicine. Finally extraction of defatted *Dryopteris filix*-mas extract was done with hydro-alcoholic solvent and % yield was found to be 6.55% w/w and their characteristics are reported. Results obtained from qualitative chemical tests are tabulated. Total alkaloid content was calculated as atropine equivalent mg/100 mg using the equation based on the calibration curve: Y=0.007X+0.007, $R^2=0.999$, where X is the Atropine equivalent (AE) and Y is the absorbance. Total phenol content was expressed as mg/100 mg of gallic acid equivalent of dry extract sample using the equation obtained from the calibration curve: Y = 0.017X+0.017, $R^2= 0.998$, where X is the gallic acid equivalent (GAE) and Y is the absorbance. The present findings indicated that the usefulness of the hydroalcoholic extract of leaves of *Dryopteris filix*-mas in earthworm. This study suggested, hydroalcoholic extract of leaves of *Dryopteris filix*-mas dose- dependently produced anthelmintic activity.

Compliance with ethical standards

Disclosure of conflict of interest

There are no conflicts of interests.

References

- [1] Parekh J, Darshana J, Sumitra C. Efficacy of Aqueous and Ethanol Extracts of some Medicinal Plants for Potential Antibacterial Activity. Turk J Biol 2005; 29: 203-210.
- [2] Kokate CK, Purohit AK, Gokhale SB. History, definition and scope of Pharmacognosy. Pharmacognosy 2005. pp. 1-14.
- [3] Kwa M.S, Veenstra, JG, Roos M H. Benzimidazole resistance in Haemonchus contortus is correlated with a conserved mutation at amino acid 200 in beta-tubulin isotype. Mol. Biochem Parasitol 1994; 63: 299–303.
- [4] Mohammed MS, Mohammed M, Yusuf OA, Joseph OA. Anthelmintic activity of the crude methanol extracts of Xylopia aethiopica against Nippostrongylus brasiliensis in rats. Veterinarski Arhiv 2005; 75: 487-495.
- [5] Geary TG, Sangster, NC, Thompson, DP. Frontiers in anthelmintic pharmacology. Vet Parasitol 1999; 84: 275–295.
- [6] Greenberg RM. Ca2+ signaling, voltage-gated Ca2+ channels and praziquantel in flatworm neuromusculature. Parasitol 2005; 131: S97–S108.
- [7] Lee BH, Clothier MF, Dutton FE, Nelson SJ, Johnson SS, Thompson DP, et al. Marcfortine and Paraherquamide class of anthelmintics: Discovery of PNU-141962. Curr. Top. Med. Chem.2002; 2: 779–793.
- [8] Schaeffer JM, Blizzard TA, Ondeyka J, Goegelman R, Sinclair PJ, Mrozik H. [3H]-Paraherquamide binding to Caenorhabditis elegans. Studies on a potent new anthelmintic agent. Biochem Pharmacol 1992; 43: 679–684.
- [9] Anisuzzaman M, Rahman AHMM, Harun-or-Rashid M, Naderuzzaman ATM, Islam AKMR an Ethnobotanical Study of Mdahupur, Tangail. J App Sci Research 2007; 3: 519-530.

- [10] Vijayan A, Liju VB, Reena John JV, Parthipan B, Renekac. Traditional remedies of kani tribes of kotor reserve forest, Agasthyavanam, Thiruvanathapuram kerala. Ind. J Trad Know 2007; 6 : 589-594.
- [11] Aceves J, Erlij D, Martinez-Maranon, R. The mechanism of the paralyzing action of tetramisole on Ascaris somatic muscle. Br J Pharmacol 1970; 38: 602–607.
- [12] Srividya A. R., Dhanabal S. P., Ajit Kumar, Y., Sathish Kumar, M. N., & Vishnuvarthan, V. J. Phytopreventive antithyperlipidemic activity of Curcuma zedoaria. Bulletin of Pharmaceutical Research, 2012; 2(1):22-25.
- [13] Ahmed, A., Howladar, S., Mohamed, H., & Al-Robai, S. Phytochemistry, Antimicrobial, Antigiardial and Antiamoebic Activities of Selected Plants from Albaha Area, Saudi Arabia. Br. J. Med. Med. Res, 2016;18(11):1-8.
- [14] Ghosh, T., Maity, T.K., Bose, A., and Dash, G.K., "Anthelmintic activity of Bacopa monierri", Indian J. Nat Prodct, 2005;21:16-19.