

## Diagnosis of thyroid malignancy using trace elements of nodular tissue determined by neutron activation analysis

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World Journal of Advanced Research and Reviews, 2022, 13(03), 438–450

Publication history: Received on 11 February 2022; revised on 22 March 2022; accepted on 24 March 2022

Article DOI: <https://doi.org/10.30574/wjarr.2022.13.3.0252>

### Abstract

Thyroid benign (TBN) and malignant (TMN) nodules are a common thyroid lesion. The differentiation of TMN often remains a clinical challenge and further improvements of TMN diagnostic accuracy are warranted. The aim of present study was to evaluate possibilities of using differences in trace elements (TEs) contents in nodular tissue for diagnosis of thyroid malignancy. Contents of TEs such as silver (Ag), cobalt (Co), chromium (Cr), iron (Fe), mercury (Hg), iodine (I), rubidium (Rb), antimony (Sb), scandium (Sc), selenium (Se), and zinc (Zn) were prospectively evaluated in “normal” thyroid (NT) of 105 individuals as well as in nodular tissue of thyroids with TBN (79 patients) and to TMN (41 patients). Measurements were performed using non-destructive instrumental neutron activation analysis. It was observed that in TMN tissue the mean mass fraction of I was lower while the mean mass fraction of Rb was higher than in NT and TBN tissue. It was demonstrated that I content in nodular tissue is the most informative parameter for the diagnosis of thyroid malignancy. It was found that “Sensitivity”, “Specificity” and “Accuracy” of TMN identification using the I level in the needle biopsy of affected thyroid tissue was significantly higher than that using US examination and cytological test of fine needle aspiration biopsy. It was concluded that determination of the I level in a needle biopsy of TNs using non-destructive instrumental analytical method is a fast, reliable, and very informative diagnostic tool that can be successfully used as an additional test of thyroid malignancy identification.

**Keywords:** Diagnosis of thyroid malignancy; Normal thyroid; Thyroid nodules; Trace elements; Neutron activation analysis

### 1. Introduction

Nodules are a common thyroid lesion, particularly in women. Depending on the method of examination and general population, thyroid nodules (TNs) have an incidence of 19–68% [1]. In clinical practice, TNs are classified into benign (TBN) and malignant (TMN), and among all TNs approximately 10% are TMN [2]. It is appropriate mention here that the incidence of TMN is increasing rapidly (about 5% each year) worldwide [2]. Surgical treatment is not always necessary for TBN whereas surgical treatment is required in TMN. Thus, differentiated TBN and TMN have a great influence on thyroid therapy.

Ultrasound (US) examination widely use as the primary method for early detection and diagnosis of the TNs. However, there are many similarities in the US characteristics of both TBN and TMN. For misdiagnosis prevention some computer-diagnosis systems based on the analysis of US images were developed, however as usual these systems for the diagnosis of TMN showed accuracy, sensitivity, and specificity nearly 80% [2,3]. Therefore, when US examination shows suspicious signs, an US-guided fine-needle aspiration biopsy is advised. Despite the fine needle aspiration biopsy has remained the diagnostic tool of choice for evaluation of US suspicious thyroid nodules, the differentiation of TMN often remains a diagnostic and clinical challenge since up to 30% of nodules are categorized as cytologically “indeterminate”

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[4]. Thus, to improve diagnostic accuracy of TMN, new technologies have to be developed for clinical applications. However, a recent systematic review and meta-analysis of molecular tests in the preoperative diagnosis of indeterminate TNs shown that at the current time there is no perfect biochemical, immunological, and genetic biomarkers to discriminate malignancy [5]. Therefore, further improvements of TMN diagnostic accuracy are warranted.

During the last decades it was demonstrated that besides the iodine deficiency and excess many other dietary, environmental, and occupational factors are associated with the TNs incidence [3,6-11]. Among these factors a disturbance of evolutionary stable input of many trace elements (TEs) in human body after industrial revolution plays a significant role in etiology of TNs [12]. Besides iodine, many other TEs have also essential physiological role and involved in thyroid functions [13]. Essential or toxic (goitrogenic, mutagenic, carcinogenic) properties of TEs depend on tissue-specific need or tolerance, respectively [13]. Excessive accumulation or an imbalance of the TEs may disturb the cell functions and may result in cellular proliferation, degeneration, death, benign or malignant transformation [13-15].

In our previous studies the complex of *in vivo* and *in vitro* nuclear analytical and related methods was developed and used for the investigation of iodine and other TEs contents in the normal and pathological thyroid [16-22]. Iodine level in the normal thyroid was investigated in relation to age, gender and some non-thyroidal diseases [23, 24]. After that, variations of many TEs content with age in the thyroid of males and females were studied and age- and gender-dependence of some TEs was observed [25-41]. Furthermore, a significant difference between some TEs contents in colloid goiter, thyroiditis, thyroid adenoma, and cancer in comparison with normal thyroid and thyroid tissue adjacent to TNs was demonstrated [42-48].

The present study had two aims. The main objective was to assess the silver (Ag), cobalt (Co), chromium (Cr), iron (Fe), mercury (Hg), iodine (I), rubidium (Rb), antimony (Sb), scandium (Sc), selenium (Se), and zinc (Zn) contents in “normal” thyroid (NT) as well as in nodular tissue of patients who had either TBN or TMN using a combination of non-destructive instrumental neutron activation analysis with high resolution spectrometry of short- and long-lived radionuclides (INAA-SLR and INAA-LLR, respectively). The second aim was to evaluate TEs content to aid diagnosis of thyroid malignancy.

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## 2. Material and methods

Samples of the NT were obtained from randomly selected autopsy specimens of 105 deceased (European-Caucasian, mean age  $44 \pm 21$  years, range 2-87), who had died suddenly. The majority of deaths were due to trauma. All the deceased were citizens of Obninsk and had undergone routine autopsy at the Forensic Medicine Department of City Hospital, Obninsk. A histological examination in the control group was used to control the age norm conformity, as well as to confirm the absence of micro-nodules and latent cancer.

All patients suffered from TBN ( $n=79$ , mean age  $M \pm SD$  was  $44 \pm 11$  years, range 22-64) and from TMN ( $n=41$ , mean age  $M \pm SD$  was  $46 \pm 15$  years, range 16-75) were hospitalized in the Head and Neck Department of the Medical Radiological Research Centre (MRRC), Obninsk. Thick-needle puncture biopsy of suspicious nodules of the thyroid was performed for every patient, to permit morphological study of thyroid tissue at these sites and to estimate their TEs contents. In all cases the diagnosis has been confirmed by clinical and morphological results obtained during studies of biopsy and resected materials. Histological conclusions for TBN were: 46 colloid goiter, 19 thyroid adenoma, 8 Hashimoto's thyroiditis, and 6 Riedel's Struma, whereas for TMN were: 25 papillary adenocarcinomas, 8 follicular adenocarcinomas, 7 solid carcinomas, and 1 reticulosarcoma. Samples of nodular tissue for INAA-SLR and INAA-LLR analysis were taken from both biopsy and resected materials.

All studies were approved by the Ethical Committees of MRRC. All the procedures performed in studies involving human participants were in accordance with the ethical standards of the institutional and/or national research committee and with the 1964 Helsinki declaration and its later amendments, or with comparable ethical standards. Informed consent was obtained from all individual participants included in the study.

All tissue samples obtained from NT, TBN and TMN were divided into two portions using a titanium scalpel to prevent contamination by TEs of stainless steel [49]. One was used for morphological study while the other was intended for TEs analysis. After the samples intended for TEs analysis were weighed, they were freeze-dried and homogenized [50].

To determine the contents of the TEs by comparison with known data for standard, aliquots of commercial, chemically pure compounds and synthetic reference materials were used [51]. In addition, ten certified reference material IAEA H-4 (animal muscle) and IAEA HH-1 (human hair) sub-samples were treated and analyzed in the same conditions that thyroid samples to estimate the precision and accuracy of results.

The content of I were determined by INAA-SLR using a horizontal channel equipped with the pneumatic rabbit system of the WWR-c research nuclear reactor (Branch of Karpov Institute, Obninsk). Details of used nuclear reaction, radionuclide, gamma-energies, spectrometric unit, sample preparation, and the quality control of results were presented in our earlier publications concerning the INAA-SLR of I contents in human thyroid [19,27,28].

A vertical channel of the same nuclear reactor was applied to determine the content of Ag, Co, Cr, Fe, Hg, Rb, Sb, Sc, Se, and Zn by INAA-LLR. Details of used nuclear reactions, radionuclides, gamma-energies, spectrometric unit, sample preparation and procedure of measurement were presented in our earlier publications concerning the INAA-LLR of TEs contents in human thyroid [29, 30] and prostate [52].

All samples for TEs analysis were prepared in duplicate, and mean values of TEs contents were used in final calculation. Using Microsoft Office Excel software, some basic statistics, including, arithmetic mean, standard deviation of mean, standard error of mean, minimum and maximum values (range) was calculated for TEs contents in three groups of thyroid tissue (NT, TBN and TMN). The difference in the results between three groups of samples was evaluated by the parametric Student's *t*-test and non-parametric Wilcoxon-Mann-Whitney *U*-test.

### 3. Results

Table 1 depicts certain statistical parameters (arithmetic mean, standard deviation, standard error of mean, range) of the Ag, Co, Cr, Fe, Hg, I, Rb, Sb, Sc, Se, and Zn mass fraction in thyroid tissue samples of three groups – NT, TBN and TMN.

**Table 1** Basic statistical parameters of Ag, Co, Cr, Fe, Hg, I, Rb, Sb, Sc, Se and Zn mass fraction (mg/kg, dry mass basis) in normal thyroid (N) and in thyroid benign (TBN) and malignant (TMN) nodules

El	NT, n=105		TBN, n=79		TMN, n=41	
	Mean± SD(SEM)	Range	Mean± SD(SEM)	Range	Mean± SD(SEM)	Range
Ag	0.015±0.014(0.0016)	0.0012-0.080	0.226±0.219(0.031)	0.0020-0.874	0.193±0.215(0.041)	0.00750-1.02
Co	0.040±0.027(0.0030)	0.0046-0.140	0.062±0.033(0.005)	0.0083-0.159	0.055±0.031(0.006)	0.0042-0.143
Cr	0.539±0.272(0.032)	0.130-1.30	0.966±0.844(0.121)	0.075-3.65	0.835±0.859(0.157)	0.0390-3.50
Fe	225±100(11)	51.0-512	332±332(40)	52.3-1407	248±173(28)	55.1-880
Hg	0.042±0.036(0.0041)	0.0065-0.180	0.924±0.649(0.088)	0.0817-3.01	0.834±0.844(0.149)	0.0685-3.75
I	1841±1027(107)	114-5061	992±901(103)	29.0-3906	71.8±62.0(10)	2.00-261
Rb	7.37±4.10(0.44)	1.11-29.4	9.55±4.37(0.52)	1.00-22.1	12.8±4.9(0.8)	5.50-27.4
Sb	0.111±0.072(0.008)	0.0047-0.308	0.137±0.116(0.016)	0.0024-0.466	0.124±0.081(0.015)	0.016-0.381
Sc	0.005±0.004(0.001)	0.0002-0.014	0.014±0.022(0.003)	0.0002-0.091	0.008±0.013(0.002)	0.0002-0.057
Se	2.32±1.29(0.14)	0.439-5.80	2.75±2.13(0.29)	0.720-12.6	2.04±1.02(0.18)	0.143-4.70
Zn	97.8±42.3(4.5)	8.10-221	118±50(6)	47.0-278	95.1±78.9(12.6)	36.5-375

El – element, M – arithmetic mean, SD – standard deviation, SEM – standard error of mean, Range – min and max values

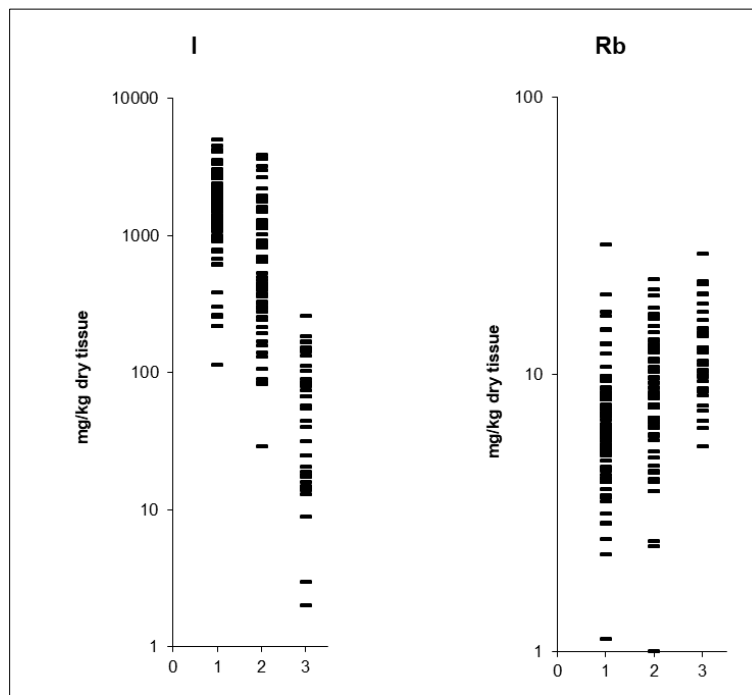
The ratios of means and the comparison of mean values of Ag, Co, Cr, Fe, Hg, I, Rb, Sb, Sc, Se, and Zn mass fractions in pair of sample groups such as NT and TBN, NT and TMN, and also TBN and TMN is presented in Table 2.

**Table 2** Ratio of means and the difference between mean values Br, Ca, Cl, I, K, Mg, Mn, and Na mass fraction (mg/kg, dry mass basis) in normal thyroid (NT) and in thyroid benign (TBN) and malignant (TMN) nodules

El	TBN and NT			TMN and NT			TMN and TBN		
	Ratio TBN/NT	<i>p</i> t-test	<i>p</i> U-test	Ratio TMN/NT	<i>p</i> t-test	<i>p</i> U-test	Ratio TMN/TBN	<i>p</i> t-test	<i>p</i> U-test
Ag	15,0	<b>&lt;0.00001</b>	<b>≤0.01</b>	12.8	<b>0.00022</b>	<b>≤0.01</b>	0.854	0.515	>0.05
Co	1,54	<b>0.00016</b>	<b>≤0.01</b>	1.38	<b>0.022</b>	<b>≤0.01</b>	0.894	0.370	>0.05
Cr	1,79	<b>0.0012</b>	<b>≤0.01</b>	1.55	0.073	>0.05	0.864	0.511	>0.05
Fe	1,48	<b>0.012</b>	<b>≤0.01</b>	1.10	0.443	>0.05	0.747	0.094	>0.05
Hg	21,9	<b>&lt;0.00001</b>	<b>≤0.01</b>	19.8	<b>0.00001</b>	<b>≤0.01</b>	0.903	0.567	>0.05
I	0.54	<b>&lt;0.0001</b>	<b>≤0.01</b>	0.039	<b>&lt;0.0001</b>	<b>≤0.01</b>	0.072	<b>&lt;0.0001</b>	<b>≤0.01</b>
Rb	1,30	<b>0.0016</b>	<b>≤0.01</b>	1.74	<b>&lt;0.00001</b>	<b>≤0.01</b>	1.340	<b>0.00098</b>	<b>≤0.01</b>
Sb	1,23	0.143	>0.05	1.12	0.423	>0.05	0.905	0.572	>0.05
Sc	3,13	<b>0.0054</b>	<b>≤0.01</b>	1.67	0.223	>0.05	0.535	0.105	>0.05
Se	1,19	0.174	>0.05	0.88	0.235	>0.05	0.742	<b>0.039</b>	<b>≤0.01</b>
Zn	1,21	<b>0.0086</b>	<b>≤0.01</b>	0.97	0.839	>0.05	0.806	0.111	>0.05

El – element, *t*-test - Student’s *t*-test, U-test - Wilcoxon-Mann-Whitney *U*-test, bold significant differences

Fig. 1 depicts individual data sets for I and Rb mass fraction in all samples of NT, TBN, and TMN group.



**Figure 1** Individual data sets for I and Rb mass fractions in samples of normal thyroid (1), thyroid benign nodules (2) and thyroid malignant nodules (3)

Parameters of the sensitivity, specificity and accuracy ( $M \pm 95\%$  confidence interval) of using I mass fraction for the diagnosis of thyroid malignancy are presented in Table 3. An estimation was made from comparison individual values in TMN group with those in NT and TBN groups combined, if value of I mass fraction equals 145 mg/kg dry tissue was chosen as upper limit (cut off) for thyroid malignancy.

**Table 3** Parameters of the sensitivity, specificity and accuracy (M±95% confidence interval) of I mass fraction for the diagnosis of TMN (an estimation is made for “TMN or NT and TBN”)

Element	Upper limit for TMN (cut off)	Sensitivity %	Specificity %	Accuracy %
I	145 mg/kg dry tissue	84±6	96±2	94±2

NT - normal thyroid, TBN - thyroid benign nodules, TMN- thyroid malignant nodules

The comparison of our results with published data (from 1990 year) for I mass fraction in NT [27,28,31-34,37,53-72], TBN [54,56,57,62,63,67-80], and TMN [54,56, 57, 60, 64-66,73,74,81-85] is shown in Tables 4, 5, and 6, respectively. A number of values for TEs mass fractions were not expressed on a dry mass basis by the authors of the cited references. However, we calculated these values using published data for water (75%) [86] And ash (4.16% on dry mass basis) [87] contents in thyroid of adults.

#### 4. Discussion

As was shown before [19, 27-30, 52] good agreement of the Ag, Co, Cr, Fe, Hg, I, Rb, Sb, Sc, Se, and Zn contents in CRM IAEA H-4 and and CRM IAEA HH-1 samples analyzed by instrumental neutron activation analysis with the certified data of these CRMs indicates acceptable accuracy of the results obtained in the study of NT, TBN, and TMN groups of tissue samples presented in Tables 1-3 and Figure 1.

From Table 2, it is observed that in TMN tissue the mass fraction of I is significantly lower while the mass fraction of Rb is higher than in NT and TBN tissue. However, as illustrated in Figure 1, I content is the most informative parameter for the diagnosis of TMN (Fig. 1). If the I level of 145 mg/kg dry tissue (about M+SD) is chosen as the upper limit (cut off) for TMN tissue (Fig.1), results for a “malignant or non- malignant” determination from results obtained would be the following:

- Sensitivity = {correct positive test (CPT)/[CPT + false negative test (FNT)]}×100% = 84±6%;
- Specificity = {correct negative test (CNT)/[CNT + false positive test (FPT)]} ×100% = 96±2%;
- Accuracy = [(CPT+CNT)/ (CPT+FNT+CNT+FPT)] ×100% = 94±2%.

The number of people examined was taken into account for calculation of confidence intervals [88]. In other words, if I contents in a nodule biopsy sample do not exceed 145 mg/kg dry tissue, one could diagnose a malignant tumor with an accuracy of 94±2%. Using the I-test makes it possible to diagnose thyroid malignancy in 84±6% cases (sensitivity).

Thus, I content in a nodule biopsy as biomarker of TMN could become a powerful diagnostic tool. To a large extent, the resumption of the search for new methods for diagnosis of TMN was due to experience gained in a critical assessment of the limited capacity of US examination and cytological test of fine needle aspiration biopsy [2-4]. In addition to the US examination and morphological study of needle-biopsy of the thyroid nodules, the I-test developed in the present study seems to be very useful. Experimental conditions of the present study were approximated to the hospital conditions as closely as possible. In all cases a part of the material obtained from a puncture needle biopsy of the affected site in the thyroid was analyzed. Therefore, our data allow us to evaluate adequately the importance of the I-test for the diagnosis of TMN. Obtained characteristics for accuracy, sensitivity, and specificity of the I-test 94, 96, and 84, respectively, are significantly better than these parameters of the US examination (nearly 80%) [2,3]. At that, the I-test gives a definite conclusion for all nodules investigated while using the morphological study of needle-biopsy up to 30% of nodules are categorized as cytologically “indeterminate” [4].

Mean values obtained for I contents in NT, TBN, and TMN agree well with median of mean values published in scientific literature for period from 1990 up to 2022 year (Table 4, 5, and 6, respectively). The range of means of I level reported in the literature for NT, TBN, and TMN vary widely (Tables 4-6). This can be explained by a dependence of I content on many factors, including age, gender, ethnicity, mass of the TNs, and the stage of diseases. Not all these factors were strictly controlled in cited studies. However, in our opinion, the leading causes of inter-observer variability can be attributed to the accuracy of the analytical techniques, sample preparation methods, and inability of taking uniform samples from the affected tissues. It was insufficient quality control of results in these studies. In many scientific reports, tissue samples were ashed or dried at high temperature for many hours. In other cases, thyroid samples were treated with solvents (distilled water, ethanol, formalin etc). There is evidence that during ashing, drying and digestion at high temperature some quantities of I are lost as a result of this treatment [89-91].

**Table 4** Reference data of I mass fractions in “normal” human thyroid published from 1990 year

Reference	Method	n	Age, years M(Range)	Sample preparation	I, mg/kg dry tissue	
					M±SD	Range
Handl et al. 1990 [53]	Chem	39	21-86	-	1276±664	-
Aeschimann et al. 1994 [54]	Chem	1	-	AD	2028	-
Boulyga et al. 1997 [55]	NAA	29	-	D, A	1778±381	-
	NAA	10	-	D, A	1905±635	-
Boulyga et al. 1999 [56]	NAA	12	-	D, A	-	800-2950
Reddy et al. 2002 [57]	PIXE	4	-	D, Press	916±88	-
Wang et al. 2002 [58]	-	21	Adult	-	2712±800	-
Murillo et al. 2005 [59]	Color	5	30-43	AD	948-3356	948-3356
Hansson et al. 2008 [60]	EDXRF	10	57-80	Intact	2400	1200-4800
Zabala et al. 2009 [61]	SFI	50	17-60	AD	5772±2708	1676-13720
Zhu et al. 2010 [62]	ICPMS	50	20-60	AD	2648	964-4760
Blázquez et al. 2011 [63]	IC	50	M=25	Fixed	601±192	624-4020
				Frozen	623±187	840 -4000
Zaichick et al. 2017 [27]	NAA	72	2-80	Intact	1786±940	220-4205
Zaichick et al. 2017 [28]	NAA	33	3.5-87	Intact	1956±1199	114-5061
Zaichick et al. 2018 [31]	EDXRF,NAA	72	2-80	Intact	1786±940	220-4205
Zaichick et al. 2018 [32]	EDXRF,NAA	33	3.5-87	Intact	1956±1199	114-5061
Zaichick et al. 2018 [33]	NAA,ICPAES	33	3.5-87	Intact	1956±1199	114-5061
Zaichick et al. 2018 [34]	NAA,ICPAES	72	2-80	Intact	1786±940	220-4205
Zaichick et al. 2018 [37]	NAA	105	2-80	Intact	1841±1027	114-5061
Zaichick et al. 2018 [64]	NAA	105	44±21	Intact	1841±1027	114-5061
Zaichick et al. 2018 [65]	NAA	105	2-80	Intact	1841±1027	114-5061
Zaichick et al. 2018 [66]	NAA	105	44±21	Intact	1841±1027	114-5061
Zaichick, 2021 [67]	NAA	105	2-87	Intact	1841±1027	114-5061
Zaichick, 2021 [68]	NAA	105	44±21	Intact	1841±1027	114-5061
Zaichick, 2021 [69]	NAA	105	2-87	Intact	1841±1027	114-5061
Zaichick, 2021 [70]	NAA	105	44±21	Intact	1841±1027	114-5061
Zaichick, 2021 [71]	NAA,ICPAES	105	2-87	Intact	1841±1027	114-5061
Zaichick, 2021 [72]	NAA,ICPAES	105	44±21	Intact	1841±1027	114-5061
Median of means	1841 mg/kg dry tissue					
Range of means (M <sub>min</sub> - M <sub>max</sub> ),	(601 - 5772) mg/kg dry tissue					
Ratio M <sub>max</sub> /M <sub>min</sub>	9.6					
All references	27					

M – Arithmetic mean, SD – standard deviation of mean; Chem – chemical method, NAA – neutron activation analysis, PIXE – proton induced X-ray fluorescent emission, Color – colorimetric method, EDXRF – energy dispersive X-ray fluorescent analysis, SFI – spectrophotometric flow injection method, ICPMS – inductively coupled plasma mass spectrometry, IC – ion chromatography, ICPAES – inductively coupled plasma atomic emission spectrometry; AD – acid digestion, D – drying at high temperature, A – ashing, AD – acid digestion

**Table 5** Reference data of I mass fractions in thyroid benign nodules published from 1990 year

Reference	Method	n	Age, years M(Range)	Sample preparation	I, mg/kg dry tissue	
					M±SD	Range
Nishita et al. 1990 [73]	NAA	14	28-71	Washed	396±74	66-1028
	NAA	7	18-74	Washed	115±40	21-344
Aeschmann et al.1994 [54]	Chem	11	-	AD	516	92-3548
Bellisola et al. 1998 [74]	NAA	20	17-82	Washed	660 ±360	560 -910
	NAA	22		Washed	1140 ±1640	7 - 3810
	NAA	12		Washed	640 ±660	3 - 1840
	NAA	6		Washed	130 ± 120	4 - 330
Boulyga et al. 1999 [56]	NAA	19	-	Washed -	-	100-4050
Reddy et al. 2002 [57]	PIXE	4	-	D, Press	888±88	-
Zhu et al. 2010 [62]	ICPMS	50	20-60	AD	2648	964-4760
Błazewicz et al. 2011 [63]	IC	50	M=25	Fixed	601±192	624-4020
	IC	50		Frozen	623±187	840 -4000
	IC	66	M=35	Fixed	77±14	41-104
Zaichick, 2021 [67]	NAA	46	30-64	Intact	1141±931	29-3715
Zaichick, 2021 [68]	NAA	19	41±11	Intact	961±1013	131-3906
Zaichick, 2021 [69]	NAA	8	40±10	Intact	951±630	83-1787
Zaichick, 2021 [70]	NAA	6	39±9	Intact	276±283	85-824
Zaichick, 2021 [71]	NAA,ICPAES	46	30-64	Intact	1141±931	29-3715
Zaichick, 2021 [72]	NAA,ICPAES	19	41±11	Intact	961±1013	131-3906
Zaichick, 2021 [75]	EDXRF,NAA	46	30-64	Intact	1144±943	29-3715
Zaichick, 2021 [76]	EDXRF,NAA	19	22-55	Intact	962±1013	131-3906
Zaichick, 2021 [77]	EDXRF,NAA	8	34-55	Intact	951±630	83-1787
Zaichick, 2021 [78]	NAA	6	34-50	Intact	276±283	85-824
Zaichick, 2022 [79]	EDXRF	79	22-64	Intact	1107±1358	47-8260
Zaichick, 2022 [80]	NAA,ICPAES	79	22-64	Intact	1086±1219	29-8260
Median of means	920 mg/kg dry tissue					
Range of means ( $M_{\min}$ - $M_{\max}$ ),	(77 - 2648) mg/kg dry tissue					
Ratio $M_{\max}/M_{\min}$	34.4					
All references	20					

M – arithmetic mean, SD – standard deviation of mean; NAA – neutron activation analysis, Chem – chemical method, PIXE – proton induced X-ray fluorescent emission, ICPMS – inductively coupled plasma mass spectrometry, IC – ion chromatography, ICPAES – inductively coupled plasma atomic emission spectrometry, EDXRF – energy dispersive X-ray fluorescent analysis; AD – acid digestion

It is well known that compared to other soft tissues, the human thyroid gland has significantly higher levels of I, because this element plays an important role in its normal functions, through the production of thyroid hormones (thyroxin and triiodothyronine) which are essential for cellular oxidation, growth, reproduction, and the activity of the central and autonomic nervous system. As was shown in present study, malignant transformation is accompanied by a significant loss of tissue-specific functional features, which leads to a drastically reduction in I content associated with functional

characteristics of the human thyroid tissue. However, it is necessary to keep in mind that biochemical, or in other words, functional changes in thyroid cells are present from the earliest development of malignancy, which precedes any histopathological indication of malignancy, and these biochemical changes persist during progression of the malignancy and remain present in advanced thyroid cancer. Thus, I depletion is an early step in the malignant proliferation process and I depletion in nodular tissue precedes the morphological transformation of cells from being histopathologically benign to malignant.

In our study non-destructive INAA-SLR was used for I determination. This method needs in using a nuclear reactor that is not always available in clinical practice. However there is an alternative non-destructive method such as EDXRF analysis, including “the total reflection” version (TRXRF), which allows reliable determinations of I and many other TES contents in a microprobe of a human body tissues and fluids within a few minutes [92]. EDXRF is a fully instrumental and non-destructive method because sample is investigated without requiring any pretreatment or its consumption. Moreover, it is well known that among the most modern analytical technologies, EDXRF is one of the simplest, fastest, most reliable and efficient of the available techniques for TES determination [92]. There are many different kinds of EDXRF and TRXRF device on the market and technical improvements are frequently announced. Thus, in our opinion, obtaining the I level in a needle biopsy of thyroid nodule, using EDXRF, is a fast, reliable and very informative diagnostic tool that can be successfully used as an additional test for diagnoses of thyroid malignancy.

**Table 6** Reference data of I mass fractions in thyroid malignant nodules published from 1990 year

Reference	Method	n	Age, years M(Range)	Sample preparation	I, mg/kg dry tissue	
					M±SD	Range
Nishida et al 1990 [73]	NAA	8	21-67	Washed	≤23±10	<DL-67
Aeschimann et al 1994 [54]	Chem	4	-	AD	40	16-140
Bellisola et al 1998 [74]	NAA	12	17-82	Washed	200±210	6 -430
Boulyga et al 1999 [56]	NAA	19	-	-	-	32-900
Reddy et al 2002 [57]	PIXE	4	-	D, Press	<30	-
Hansson et al 2008 [60]	EDXRF	7	21-58	Intact	<400	-
Zaichick et al. 2018 [64]	NAA	41	16-75	Intact	71.8±62	2-261
Zaichick et al. 2018 [65]	EDXRF,NAA	41	46±15	Intact	71.8±62	2-261
Zaichick et al. 2018 [66]	NAA,ICPAES	41	16-75	Intact	71.8±62	2-261
Zaichick, 2022 [81]	EDXRF	41	16-75	Intact	71.6±72.5	2-341
Zaichick, 2022 [82]	NAA	41	16-75	Intact	71.8±62	2-261
Zaichick, 2022 [83]	NAA	41	16-75	Intact	71.8±62	2-261
Zaichick, 2022 [84]	EDXRF,NAA	41	16-75	Intact	71.8±62	2-261
Zaichick, 2022 [85]	NAA,ICPAES	41	16-75	Intact	71.8±62	2-261
Median of means	71.8 mg/kg dry tissue					
Range of means ( $M_{\min}$ - $M_{\max}$ ),	(23 - 400) mg/kg dry tissue					
Ratio $M_{\max}/M_{\min}$	17.4					
All references	14					

M - arithmetic mean, SD - standard deviation of mean; NAA - neutron activation analysis, Chem - chemical method, PIXE - proton induced X-ray fluorescent emission, EDXRF - energy dispersive X-ray fluorescent analysis, ICPAES - inductively coupled plasma atomic emission spectrometry; AD - acid digestion, D - drying at high temperature

## 5. Conclusion

In this work, TES analysis was carried out in the tissue samples of NT and thyroid with TBN and TMN using two methods of neutron activation analysis - INAA-SLR and INAA-LLR. It was shown that neutron activation analysis is an adequate



analytical tool for the non-destructive determination of Ag, Co, Cr, Fe, Hg, I, Rb, Sb, Sc, Se, and Zn content in the tissue samples of human thyroid, including needle-biopsy material. It was observed that in TMN tissue the mean mass fraction of I was lower while the mean mass fraction of Rb was higher than in both NT and TBN groups of samples. It was demonstrated that I content in nodular tissue is the most informative parameter for the diagnosis of thyroid malignancy. It was found that “Sensitivity”, “Specificity” and “Accuracy” of TMN identification using the I level in the needle biopsy of affected thyroid tissue was significantly higher than that using US examination and cytological test of fine needle aspiration biopsy. It was concluded that determination of the I level in a needle biopsy of TNs, using non-destructive instrumental analytical method, is a fast, reliable, and very informative diagnostic tool that can be successfully used as an additional test of thyroid malignancy identification.

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## Compliance with ethical standards

### *Acknowledgments*

The author is extremely grateful to Profs. Vtyurin BM and Medvedev VS, Medical Radiological Research Center, Obninsk, as well as to Dr. Choporov Yu, former Head of the Forensic Medicine Department of City Hospital, Obninsk, for supplying thyroid samples.

### *Disclosure of conflict of interest*

The author declares that he has no competing interests.

### *Statement of ethical approval*

All studies were approved by the Ethical Committees of the Medical Radiological Research Centre (MRRC), Obninsk. All the procedures performed in studies involving human participants were in accordance with the ethical standards of the institutional and/or national research committee and with the 1964 Helsinki declaration and its later amendments, or with comparable ethical standards.

### *Statement of informed consent*

Informed consent was obtained from all individual participants included in the study.

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